

## Meta-Analysis

## Asian Pacific Journal of Tropical Medicine

doi: 10.4103/1995-7645.329009

5-Year Impact Factor: 2.285

## Tick-borne pathogens in Iran: A meta-analysis

Mehdi Khoobdel<sup>1</sup>, Amir Sajad Jafari<sup>2</sup>, Zakkyeh Telmadarraiy<sup>3</sup>, Mohammad Mehdi Sedaghat<sup>4</sup>, Hasan Bakhshi<sup>5</sup>✉<sup>1</sup>Health Research Center, LifeStyle Institute, Baqiyatallah University of Medical Sciences, Tehran, Iran<sup>2</sup>Department of Veterinary Medicine, Faculty of Veterinary Medicine, University of Zabol, Zabol, Iran<sup>3</sup>Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran; Rahyan Novin Danesh (RND) University, Sari, Mazandaran, Iran<sup>4</sup>Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran<sup>5</sup>Vector-borne Diseases Research Center, North Khorasan University of Medical Sciences, Bojnurd, Iran

## ABSTRACT

**Objective:** Different studies have been performed on the prevalence of tick-borne pathogens in different areas of Iran; however, as far as our knowledge, there is no regional meta-analysis available for consideration and estimation of tick species infected with different pathogens in Iran.

**Methods:** In this review, among different databases, a total of 95 publications were included, and the infection of different tick species to different tick-borne pathogens was determined; furthermore, presence of pathogens (with 95% confidence intervals) in tick vectors was calculated separately for each province, using Comprehensive Meta-Analysis version 2 (Biostat, USA).

**Results:** Totally, among all 95 studies, 5 673 out of 33 521 investigated ticks were positive according to different detection methods. Overall estimated presence of pathogens in tick vectors in Iran was 8.6% (95% CI 7.0%-10.6%,  $P < 0.001$ ). Of all 46 species of ticks in 10 genera in Iran, 28 species in 9 genera, including *Alveonassus*, *Argas*, *Boophilus*, *Dermacentor*, *Haemaphysalis*, *Hyalomma*, *Ixodes*, *Ornithodoros*, and *Rhipicephalus* were infected with at least 20 pathogens in 10 genera including *Aegyptianella*, *Anaplasma*, *Babesia*, *Borrelia*, *Brucella*, *Orthonairovirus* [Crimean-Congo hemorrhagic fever virus (CCHFV)], *Coxiella*, *Ehrlichia*, *Rickettsia* and *Theileria* in 26 provinces of Iran. The presence of pathogens in ticks collected in western Iran was more than other regions. *Hyalomma anatolicum* (20.35%), *Rhipicephalus sanguineus* (15.00%), and *Rhipicephalus bursa* (14.08%) were the most prevalent infected ticks for different pathogens. In addition, most literatures were related to CCHFV and *Theileria/Babesia* spp.

**Conclusions:** Public health and veterinary professionals should be aware of diagnosing possible diseases or outbreaks in vertebrates.

**KEYWORDS:** Ticks; Tick-borne diseases; Vector-borne diseases; Iran

## 1. Introduction

Ticks are external obligatory blood-sucking parasites of vertebrates (phylum Arthropoda; class Arachnida) that fall into three families including Ixodidae (hard ticks), Argasidae (soft ticks), and Nuttalliellidae[1]. Ticks are the primary vectors and reservoirs for different pathogens including viruses, bacteria, and protozoa all over the world, which pose significant threats to human and animal health[2,3]. Tick-borne pathogens cause thousands of disease cases in human populations worldwide with the animal cases seeming to be more than humans[4]. Different species of ticks are able to transmit different diseases. And Crimean-Congo hemorrhagic fever (CCHF), Colorado tick fever, Q fever, borreliosis, relapsing fever, theileriosis, babesiosis, anaplasmosis, ehrlichiosis and Rocky Mountain spotted fever are

## Significance

Several studies have shown the presence of tick-borne pathogens in ticks in Iran; however, as far as our knowledge, there is no meta-analysis available for estimation of ticks infected with tick-borne pathogens. Our analysis showed that the overall estimated presence of pathogens in tick vectors in Iran was 8.6% (95% CI 7.0%-10.6%,  $P < 0.001$ ). Furthermore, 28 tick species in 9 genera were found to be infected with at least 20 pathogens in 10 genera.

✉To whom correspondence may be addressed. E-mail: hbakhshi89@gmail.com

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non Commercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

**For reprints contact:** reprints@medknow.com

©2021 Asian Pacific Journal of Tropical Medicine Produced by Wolters Kluwer-Medknow. All rights reserved.

**How to cite this article:** Khoobdel M, Jafari AS, Telmadarraiy Z, Sedaghat MM, Bakhshi H. Tick-borne pathogens in Iran: A meta-analysis. Asian Pac J Trop Med 2021; 14(11): 486-504.

**Article history:** Received 15 June 2021

Revision 21 October 2021

Accepted 22 November 2021

Available online 30 November 2021

among the most significant tick-borne diseases caused by these pathogens[5]. The spectrum of tick-borne diseases of both medical and veterinary importance has increased in recent years as a result of advances in molecular biology. New microorganisms are being detected in ticks collected in different countries, and the list of potential tick-transmissible pathogens is updating[6]. Problems caused by tick infestations are not limited only to transmission of pathogens. Bite stress, production loss, physical damage, anemia and poisoning are other aspects of tick bites[7]. Furthermore, the importance of animal productions in the economy and food industry around the world is undeniable[8]. Animal health can be altered by the direct and indirect effects caused by the bites of ticks and tick-borne diseases, leading to noteworthy production decrement of meat, milk, eggs, and leathers. In some severe cases, tick-borne pathogens lead to the death of humans and animals. Indirect effects are related to the costs associated to the treatment and control[8]. From past to present, ticks and tick-borne diseases have been recognized as a threat for human and animal health. Ticks are responsible for the majority of vector-borne diseases in Asia, America and Europe[9].

Iran, covering an area of 1 648 195 km<sup>2</sup>, with a population of 83 million, is located in the Middle East. This country is located in Palearctic and Oriental zoogeographic regions, with different types of climate: mild and quite wet on the coast of the Caspian Sea, continental and arid in the plateau, cold in high mountains, desert and hot on the southern coast and in the southeast, resulting in diversity of tick species[10,11]. Ecology of ticks, their interactions with environment and risk of infection by tick-borne pathogens are directly related to the spatial and temporal variations. As a result, diversity of climate, as well as the vast geographical area, increases the diversity of tick populations which leads to the risk of transmission of different tick-borne pathogens[12]. To date, it has been reported that 46 species of ticks (10 Argasidae and 36 Ixodidae) in 10 genera occur in the country[13].

Tick species can be considered as sentinels to track the circulation of tick-borne pathogens before an outbreak breaks out in humans and animals. Although many studies revealed data about prevalence of different tick-borne pathogens in different areas of Iran, as far as our knowledge, there is no comprehensive data available for consideration and estimation of the damages caused by pathogens transmitted by ticks, on the economy and public health in Iran. For this reason, performing an updated regional review and meta-analysis on the studies conducted on the prevalence of tick-borne pathogens in different provinces of this country is highly necessary. Considering the damages caused by tick-borne diseases on the public health, animal husbandry, and Iran tourism industry, the current study attempted to determine and highlight the presence of pathogens in tick vectors and epidemiological aspects of tick-borne diseases in Iran.

## 2. Materials and methods

### 2.1. Searching approach

The present meta-analysis was performed according to the guidelines of preferred reporting items for systematic reviews and meta-analyses statement. In this regional meta-analysis study, nine English and Persian language databases including PubMed, Google Scholar, Science Direct, Scopus, Web of Science, Magiran, Civilica, Iranian Research Institute for Information Science and Technology (IranDoc), and Scientific Information Database (SID) were selected to explore the articles and data with no time limitation (last updated: 7 March, 2021). Duplicate articles, case series, animal-based studies, human-based studies and studies carried out in other countries were excluded. All studies, representing the prevalence of tick-borne pathogens in ticks as hosts/reservoirs were concerned and all PRISMA criteria have been met (Figure 1).

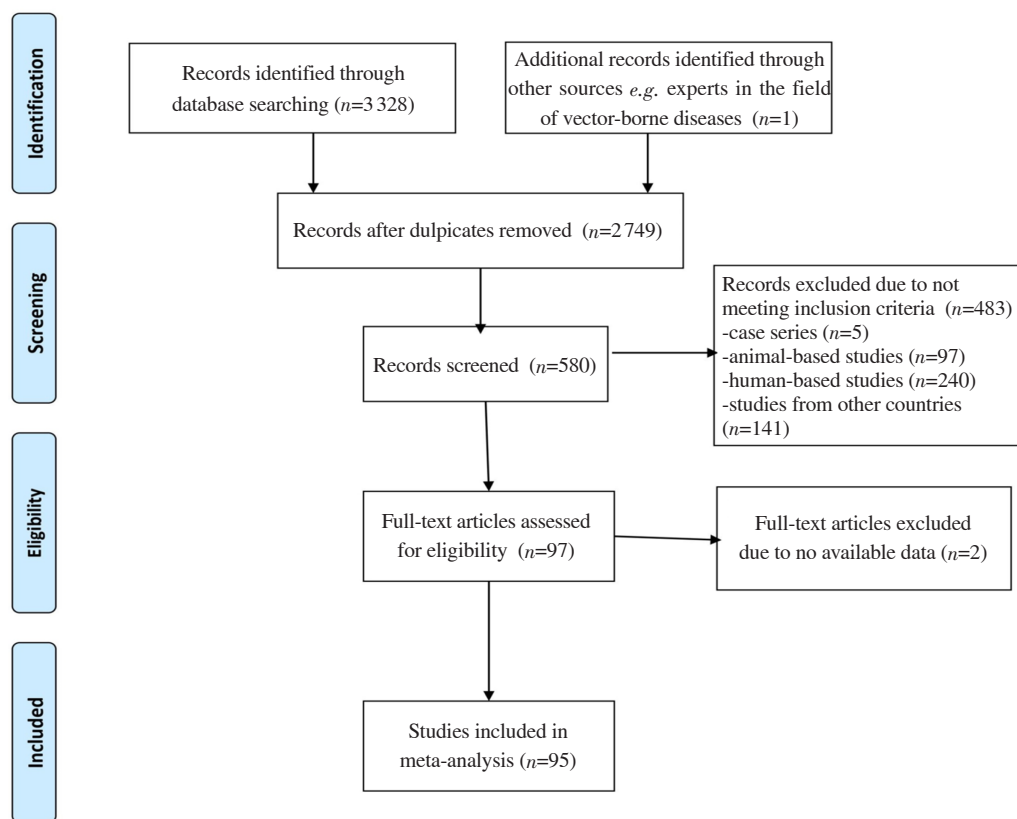
Totally, 95 articles and data fit into the criteria. Then, author(s) names, year of publication, province of study, tick vectors, pathogens, the number of examined ticks and the number of positive ticks were extracted from the collected data. The search was conducted using English and Persian language keywords with different patterns (*e.g.*: Tick, Iran, *Anaplasma*, *Babesia*, *Theileria*, Crimean-Congo hemorrhagic fever virus, CCHFV, *Ehrlichia*, *Agyptinella*, *Francisella*, *Brucella*, *Borrelia*, *Coxiella*, and *Rickettsia*). Advanced search options and Boolean operators 'AND' and 'OR' were also used to find more relevant records.

### 2.2. Paper selection

PICO process or framework (Population, Intervention, Comparator and Outcome) is a common method for formulating a systematic review queries. However, this format is not suitable for prevalence studies. Quality assessment for the included studies of the present research were setup and developed according to CoCoPop structure [Co (Condition)=infection by pathogens; Co (Context)=provinces of Iran; Pop (Population)=ticks]. Studies and the selected data were independently analyzed and the eligibility was determined by HB and ASJ. Disagreements were resolved by MK.

### 2.3. Meta-analysis

Initially, the prevalence of each genus of pathogen (with 95% confidence intervals) was calculated separately for each province (at least two studies were needed for calculation of each pathogen in separate provinces). Then, an overall prevalence was calculated for all pathogens in respect to each province. Furthermore, the total



**Figure 1.** Flowchart of studies selection in terms of tick-borne pathogens in Iran.

prevalence for each pathogen in Iran was estimated. Cochran  $Q$  test ( $P < 0.05$  shows statistically significant heterogeneity) and  $I^2$  test [25% (low), 50% (moderate), and 75% (high) heterogeneity] were used to evaluate heterogeneity among studies. To compute overall size effect ( $Q < 0.05$ ), random model was used; otherwise ( $Q > 0.05$ ), fixed model was assessed. For determination of publication bias, Egger's and Begg's tests were applied ( $P > 0.05$  indicates a reasonable publication bias). Also, a funnel plot was used to visualize the publication bias.  $P < 0.05$  was considered statistically significant for statistical analysis of prevalence. All statistical analyses were performed using Comprehensive Meta-Analysis version 2 (Biostat, USA).

### 3. Results

Among all databases screened, 3328 records were identified through database searching; then, a total of 95 publications were selected and included in this review. Among these 95 publications, 33521 ticks were surveyed and 5673 were positive according to different detection methods in all provinces of Iran. Of all 46 species of ticks (in 10 genera) which occur in Iran [13], 28 species (in 9 genera) including *Alveonassus* (1 species: *Al. canestrinii*), *Argas* (2 species: *Ar. persicus*, *Ar. reflexus*), *Boophilus* (*Boophilus* spp.), *Dermacentor* (2 species: *D. marginatus*, *D. niveus*), *Haemaphysalis*

(4 species: *Ha. concinna*, *Ha. inermis*, *Ha. punctata*, *Ha. sulcata*), *Hyalomma* (10 species: *H. aegyptium*, *H. anatolicum*, *H. asiaticum*, *H. detritum*, *H. dromedarii*, *H. excavatum*, *H. marginatum*, *H. rufipes*, *H. schulzei*, *H. scupense*, *H. sp.*), *Ixodes* (1 species: *I. ricinus*), *Ornithodoros* (3 species: *O. erraticus*, *O. lahorensis*, *O. tholozani*), and *Rhipicephalus* (5 species: *R. annulatus*, *R. appendiculatus*, *R. bursa*, *R. sanguineus*, *R. turanicus*, *R. spp.*) were found to be infected with at least 20 pathogens (in 10 genera) including *Aegyptianella* (1 species: *Ae. pullorum*), *Anaplasma* (4 species: *An. ovis*, *An. bovis*, *An. phagocytophilum*, *An. marginale*, *An. spp.*), *Babesia* (3 species: *Ba. ovis*, *Ba. bigemina*, *Ba. occultans*, *Ba. spp.*), *Borrelia* (3 species: *Bo. microti*, *Bo. anserina*, *Bo. persica*, *Bo. sp.*), *Brucella* (*Brucella* sp.), *Orthonairovirus* (1 virus: CCHFV), *Coxiella* (1 species: *Cx. burnetii*), *Ehrlichia* (2 species: *Eh. canis*, *Eh. ovina*, *Eh. spp.*), *Rickettsia* (1 species: *Ri. hoogstraalii*, *Ri. sp.*), *Theileria* (4 species: *Th. annulata*, *Th. lestoquardi*, *Th. ovis*, *Th. equi*, *Th. spp.*), as well as unspecified *An. centrale*/*An. bovis* (Table 1). In this review, *D. marginatus*, *D. niveus*, *H. detritum* and *H. scupense* were considered as separate species.

Among the provinces where ticks were found to be infected with different genera of pathogens (including CCHFV), Lorestan (7 genera), Ardabil (6 genera), Golestan (5 genera), and Sistan and Baluchestan (5 genera) provinces had the most number of ticks infected with different genera of pathogens (Table 2).

Among 31 provinces of Iran, 26 provinces were surveyed in

















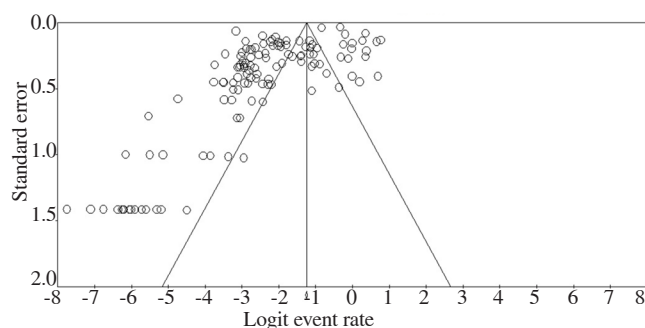
**Table 2.** Different genera of pathogens (as well as CCHFV) detected in tick vectors in different provinces of Iran.

Province	Total tested/positive tick(s)	Positive tick vector(s)	Pathogen(s)
Ardabil	1062/226	<i>D. marginatus</i> ; <i>D. niveus</i> ; <i>H. aegyptium</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. excavatum</i> ; <i>H. marginatum</i> ; <i>H. schulzei</i> ; <i>H. sp.</i> ; <i>O. lahorensis</i> ; <i>O. tholozani</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i>	<i>Babesia</i> ; <i>Borrelia</i> ; CCHFV; <i>Coxiella</i> ; <i>Ehrlichia</i> ; <i>Theileria</i>
Azerbaijan, East	998/202	<i>D. marginatus</i> ; <i>Ha. sulcata</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. marginatum</i> ; <i>I. ricinus</i> ; <i>O. lahorensis</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i>	<i>Anaplasma</i> ; <i>Babesia</i> ; <i>Brucella</i> ; CCHFV
Azerbaijan, West	1904/192	<i>D. marginatus</i> ; <i>H. marginatum</i> ; <i>R. annulatus</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i> ; <i>R. turanicus</i>	<i>Babesia</i> ; <i>Theileria</i>
Fars	550/110	<i>H. anatolicum</i> ; <i>H. marginatum</i> ; <i>H. sp.</i> ; <i>R. sanguineus</i> ; <i>R. sp.</i> ; <i>R. turanicus</i>	<i>Babesia</i> ; <i>Ehrlichia</i> ; CCHFV; <i>Theileria</i>
Gilan	591/29	<i>Boophilus</i> spp.; <i>D. marginatus</i> ; <i>I. ricinus</i> ; <i>R. annulatus</i> ; <i>R. sanguineus</i> ; <i>R. sp.</i>	<i>Anaplasma</i> ; <i>Borrelia</i> ; <i>Brucella</i>
Golestan	685/78	<i>H. anatolicum</i> ; <i>H. dromedarii</i> ; <i>H. excavatum</i> ; <i>H. marginatum</i> ; <i>H. rufipes</i> ; <i>H. scupense</i> ; <i>I. ricinus</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i> ; <i>R. turanicus</i>	<i>Anaplasma</i> ; <i>Babesia</i> ; <i>Borrelia</i> ; CCHFV; <i>Theileria</i>
Hamadan	1755/83	<i>Ar. reflexus</i> ; <i>Ha. punctata</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. dromedarii</i> ; <i>H. marginatum</i> ; <i>O. tholozani</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i>	<i>Borrelia</i> ; CCHFV
Hormozgan	30/1	<i>H. dromedarii</i>	<i>Anaplasma</i>
Ilam	137/9	NA	CCHFV
Isfahan	210/11	<i>Ha. sulcata</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. sp.</i> ; <i>R. sanguineus</i>	CCHFV
Kerman	1796/113	<i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. marginatum</i> ; <i>R. sanguineus</i>	<i>Anaplasma</i> ; <i>Coxiella</i> ; <i>Ehrlichia</i>
Kermanshah	551/135	<i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. marginatum</i> ; <i>R. sanguineus</i> ; <i>R. turanicus</i>	CCHFV; <i>Theileria</i>
Khorasan, North	497/30	<i>H. anatolicum</i> ; <i>H. marginatum</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i> ; <i>R. turanicus</i>	<i>Babesia</i> ; CCHFV; <i>Theileria</i>
Khorasan, Razavi	2707/344	<i>H. asiaticum</i> ; <i>H. excavatum</i> ; <i>H. marginatum</i> ; <i>O. tholozani</i> ; <i>R. appendiculatus</i> ; <i>R. turanicus</i>	<i>Borrelia</i> ; CCHFV; <i>Theileria</i>
Khorasan, South	553/101	<i>Ar. persicus</i> ; <i>D. niveus</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. dromedarii</i> ; <i>H. marginatum</i> ; <i>O. lahorensis</i> ; <i>R. sanguineus</i> ; <i>R. sp.</i>	<i>Anaplasma</i> ; CCHFV
Khuzestan	655/67	<i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. dromedarii</i>	<i>Theileria</i>
Kohgiluyeh and Boyer-Ahmad	469/1	<i>R. bursa</i>	CCHFV
Kurdistan	3393/2269	<i>Ha. punctata</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. dromedarii</i> ; <i>H. excavatum</i> ; <i>H. marginatum</i> ; <i>O. tholozani</i> ; <i>R. annulatus</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i>	<i>Babesia</i> ; <i>Borrelia</i> ; CCHFV; <i>Theileria</i>
Lorestan	1543/359	<i>Al. canestrinii</i> ; <i>Ar. persicus</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. marginatum</i> ; <i>R. sanguineus</i>	<i>Aegyptianella</i> ; <i>Anaplasma</i> ; <i>Borrelia</i> ; <i>Coxiella</i> ; CCHFV; <i>Rickettsia</i> ; <i>Theileria</i>
Mazandaran	2137/465	<i>D. marginatus</i> ; <i>Ha. concinna</i> ; <i>Ha. inermis</i> ; <i>Ha. punctata</i> ; <i>H. anatolicum</i> ; <i>H. dromedarii</i> ; <i>H. marginatum</i> ; <i>I. ricinus</i> ; <i>R. annulatus</i> ; <i>R. bursa</i> ; <i>R. sanguineus</i> ; <i>R. turanicus</i>	<i>Anaplasma</i> ; <i>Borrelia</i> ; CCHFV; <i>Theileria</i>
Qazvin	599/51	<i>O. erraticus</i> ; <i>O. lahorensis</i> ; <i>O. tholozani</i>	<i>Borrelia</i>
Qom	88/6	<i>H. marginatum</i>	CCHFV
Semnan	6031/247	<i>H. anatolicum</i> ; <i>H. dromedarii</i> ; <i>O. tholozani</i> ; <i>R. sanguineus</i>	<i>Borrelia</i> ; CCHFV
Sistan and Baluchestan	3533/424	<i>D. marginatus</i> ; <i>Ha. inermis</i> ; <i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. dromedarii</i> ; <i>H. excavatum</i> ; <i>H. marginatum</i> ; <i>H. schulzei</i> ; <i>H. sp.</i> ; <i>R. sanguineus</i> ; <i>R. sp.</i> ; <i>R. turanicus</i>	<i>Anaplasma</i> ; <i>Coxiella</i> ; <i>Ehrlichia</i> ; CCHFV; <i>Theileria</i>
Tehran	116/18	<i>R. sanguineus</i>	<i>Theileria</i>
Yazd	390/23	<i>H. anatolicum</i> ; <i>H. asiaticum</i> ; <i>H. detritum</i> ; <i>H. dromedarii</i> ; <i>H. marginatum</i>	CCHFV; <i>Theileria</i>

*Babesia* (7 provinces), *Borrelia* (10 provinces), CCHFV (19 provinces), *Coxiella* (4 provinces), *Ehrlichia* (4 provinces), *Theileria* (14 provinces), *Anaplasma* (9 provinces), *Brucella* (2 provinces), *Aegyptianella* (1 province), *Rickettsia* (1 province); Positive tick species in different provinces are as follows: *Al. canestrinii* (1 province), *Ar. persicus* (2 provinces), *Ar. reflexus* (1 province), *D. marginatus* (6 provinces), *D. niveus* (2 provinces), *H. aegyptium* (1 province), *H. anatolicum* (17 provinces), *H. asiaticum* (13 provinces), *H. detritum* (7 provinces), *H. dromedarii* (10 provinces), *H. excavatum* (5 provinces), *H. marginatum* (17 provinces), *H. rufipes* (1 province), *H. schulzei* (2 provinces), *H. scupense* (1 province), *Ha. concinna* (1 province), *Ha. inermis* (2 provinces), *Ha. punctata* (3 provinces), *Ha. sulcata* (2 provinces), *I. ricinus* (4 provinces), *O. erraticus* (1 province), *O. lahorensis* (4 provinces), *O. tholozani* (6 provinces), *R. annulatus* (4 provinces), *R. appendiculatus* (1 province), *R. bursa* (9 provinces), *R. sanguineus* (18 provinces), *R. turanicus* (8 provinces), *Boophilus* spp. (1 province), *Hyalomma* spp. (4 provinces), *Rhipicephalus* sp. (4 provinces), NA (1 province).

terms of detection of infection of different pathogens in ticks; meanwhile, the status of tick infection with different pathogens remained unclear in Alborz, Bushehr, Chaharmahal and Bakhtiari, Markazi, and Zanjan provinces. The provinces in which the most studies have been carried out are Sistan and Baluchestan (12 studies), Lorestan (9 studies), Razavi Khorasan (8 studies), Mazandaran (8 studies), Kerman (7 studies), and Ardabil (7 studies). On the other hand, Hormozgan, Ilam, Isfahan, Khuzestan, Kohgiluyeh and Boyer-Ahmad, and Qom were among the least studied provinces (only one study in each province). More than 60 literatures were related to CCHFV and *Theileria/Babesia* spp., while *Aegyptianella*, *Brucella* and *Rickettsia* were limited to less than 10 publications (Table 1). According to a random effect model, the total prevalence of tick-borne pathogens in Iran was calculated as 8.6% (95% CI 7.0%-10.6%,  $P<0.001$ ). The highest and lowest prevalence rate occurred in Kurdistan (20.5%; 95% CI 14.0%-29.1%,  $P<0.001$ ), and Khorasan, Razavi (2.4%; 95% CI 0.8%-6.7%,  $P=0.008$ ), respectively. In addition, *Anaplasma* sp. was the pathogen with the highest statistically significant prevalence (23.5%; 95% CI 15.1%-34.7%,  $P<0.001$ ), while the lowest infection rate belonged to *Babesia* sp. (4.0%; 95% CI 1.9%-8.1%,  $P<0.001$ ) (Table 3).

Statistical analysis revealed that the highest prevalence of *Anaplasma* sp., *Babesia* sp., *Borrelia* sp., CCHFV, *Coxiella* sp., and *Theileria* sp. occurred in East-Azerbaijan (36.5%; 95% CI 15%-63.9%,  $P=0.335$ ), West-Azerbaijan (8.8%; 95% CI 6.1%-12.5%,  $P<0.001$ ), Kurdistan (8.5%; 95% CI 1.2%-41.6%,  $P=0.022$ ), South-Khorasan (14.3%; 95% CI 3.7%-42.0%,  $P=0.017$ ), Kerman (9.9%; 95% CI 5.8%-16.4%,  $P<0.001$ ), and Mazandaran (21.0%; 95% CI 1.5%-82.4%,  $P=0.009$ ), respectively. *Brucella* sp., *Ehrlichia* sp., *Rickettsia* sp., and *Aegyptianella* sp. did not meet the criteria for entering province-specific meta-analysis (less than 2 publications in each province). A forest plot was used to show the prevalence of tick-borne pathogens across the country (Supplementary Figure 1). In addition, funnel plot revealed an asymmetry in the funnel which might indicate that some studies were missed on the right side of the plot (Figure 2). In line with funnel plot, the results of Egger's test ( $P<0.001$ ) showed a publication bias among studies. Based on the funnel plot, most of the studies with low prevalence of tick-borne pathogens were included in this meta-analysis (Figure 2).



**Figure 2.** Funnel plot of standard error by logit event rate.

## 4. Discussion

As far as we know, the present meta-analysis is the first large-scale study that examined the prevalence of tick-borne pathogens in tick vectors in Iran. Overall estimated prevalence of tick-borne pathogens in Iran was 8.6% (95% CI 7.0%-10.6%,  $P<0.001$ ). The greatest infection rates among tick vectors were dedicated to *Rickettsia* sp. ( $P>0.05$ ), and *Anaplasma* sp., respectively. *Anaplasma* species are Gram-negative obligate intraerythrocytic bacteria (Rickettsiales; Anaplasmataceae) which are of great veterinary concern. *An. marginale*, the most probable causative agent of bovine anaplasmosis, has been reported worldwide. This pathogen mainly affects ruminants resulting in mild to severe febrile hemolytic anemia that leads to significant economic losses[109]. Other species are as follows *An. ovis* and *An. mesaeterum* (in sheep and goat), *An. phagocytophilum* (in horse, dogs and cats), *An. platys* (in dogs) and *An. centrale* in cattle[110,111]. Although medically important pathogens such as *Borrelia* sp., *Coxiella* sp., and CCHFV were less prevalent in ticks according to the pooled results of literature review, it should be noted that to determine the epidemiological status of a pathogen, all factors affecting pathogen's life cycle must be taken into consideration. For example, CCHF is endemic in Iran and its neighboring countries and a significant number of human cases are reported each year. In a recent review on distribution of ticks and their infection to CCHFV, the main vectors of CCHF, *H. marginatum* and *H. anatolicum*, have been reported in more than 38.7% of provinces of Iran[112]. In our review, among all pathogens, CCHFV positive ticks were reported in 19 provinces. The point may be that in Iran, the main way of CCHFV transmission is not tick bite. CCHFV infection in human mostly occurs due to direct contact with infected livestock (blood, tissues, secretions), which have been infected by ticks[113,114].

Q fever is a zoonosis caused by the bacterium *Cx. burnetii*. Human infection mainly occurs through inhalation of contaminated animal products, direct contact with infected animals and consumption of unpasteurized milk or other dairy products contaminated with this pathogen. Ticks play a key role in transmitting bacteria between animals, and are considered as reservoirs of *Cx. burnetii* bacteria and guarantee the long-term presence of this microorganism in nature[84]. *Borrelia* spp. is the causative agent of Lyme disease and relapsing fever which are zoonotic vector-borne diseases transmitted primarily by ticks[115]. In a descriptive and retrospective study during 1997-2006, Masoumi *et al.* reported that the disease is detected in humans in 18 provinces of the 31 provinces in Iran[116]. Other reports also revealed that *Borrelia* spp. is present in ticks and other vertebrates[35,117]. According to reports of *Cx. burnetii* and *Borrelia* spp. in ticks, humans, and animals in Iran, Q-fever, Lyme disease and relapsing fever can be considered as emerging diseases in the country[118-120].

The most infected provinces in terms of tick-borne pathogens

**Table 3.** Meta-analysis result of different genera of pathogens (including CCHFV), detected in each province as well as in the country.

Provinces	Pathogens	No. of studies	Sample size	Prevalence (pooled effect size)	95% CI		Heterogeneity		P values of prevalence	Publication bias	
					Lower	Upper	I <sup>2</sup> (%)	Q test		Begg's test (2 tailed P value)	Egger's test (2 tailed P value)
Ardabil	<i>Babesia</i> sp.	2	489	0.012	0.000	0.255	81.938	5.537	0.009	NA	NA
	Tick borne pathogens	7	1351	0.186	0.104	0.310	93.410	91.046	<0.001	0.763	0.357
Azerbaijan, East	<i>Anaplasma</i> sp.	3	413	0.365	0.158	0.639	95.564	45.088	0.335	1.000	0.678
	<i>Theileria</i> sp.	2	275	0.032	0.013	0.077	71.037	3.45	<0.001	NA	NA
	Tick borne pathogens	7	998	0.133	0.051	0.305	96.091	153.502	<0.001	0.367	0.051
Azerbaijan, West	<i>Babesia</i> sp.	3	1589	0.088	0.061	0.125	75.961	8.320	<0.001	0.296	0.032
	Tick borne pathogens	4	1904	0.097	0.074	0.125	67.142	9.130	<0.001	0.308	0.176
Fars	<i>Theileria</i> sp.	3	350	0.123	0.009	0.682	97.645	84.928	0.158	1.000	0.081
	Tick borne pathogens	6	810	0.115	0.035	0.316	96.359	137.321	<0.001	0.259	0.045
Gilan	<i>Anaplasma</i> sp.	2	83	0.169	0.008	0.830	89.515	9.537	0.326	NA	NA
	Tick borne pathogens	3	590	0.049	0.001	0.699	96.472	56.697	0.127	1.000	0.334
Golestan	<i>Babesia</i> sp.	2	96	0.033	0.011	0.097	0.000	0.331	<0.001	NA	NA
	Tick borne pathogens	6	829	0.066	0.030	0.139	87.992	41.639	<0.001	0.707	0.683
Hamedan	<i>Borrelia</i> sp.	2	1239	0.005	0.000	0.278	88.288	8.538	0.017	NA	NA
	CCHFV	3	516	0.124	0.066	0.221	78.850	9.456	<0.001	0.296	0.126
	Tick borne pathogens	5	1755	0.060	0.023	0.149	88.583	35.034	<0.001	0.027	0.001
Hormozgan						NA					
Ilam						NA					
Isfahan						NA					
Kerman	<i>Coxiella</i> sp.	2	620	0.099	0.058	0.164	75.927	4.154	<0.001	NA	NA
	CCHFV	2	461	0.002	0.000	0.015	0.000	0.014	<0.001	NA	NA
	Tick borne pathogens	7	1796	0.060	0.029	0.119	88.637	88.637	<0.001	0.763	0.110
Kermanshah	<i>Theileria</i> sp.	3	1260	0.096	0.055	0.164	90.249	20.511	<0.001	0.296	0.042
	Tick borne pathogens	4	1391	0.082	0.047	0.139	88.747	26.659	<0.001	0.308	0.164
Khorasan, North	<i>Babesia</i> sp.	3	125	0.029	0.009	0.087	0.000	1.209	<0.001	1.000	0.050
	CCHFV	2	196	0.023	0.001	0.319	77.002	4.348	0.014	NA	NA
	<i>Theileria</i> sp.	4	697	0.036	0.024	0.054	37.422	4.794	<0.001	1.000	0.938
	Tick borne pathogens	9	1018	0.038	0.027	0.054	31.263	11.639	<0.001	0.676	0.219
Khorasan, Razavi	<i>Babesia</i> sp.	2	675	0.002	0.000	0.014	0.000	0.000	<0.001	NA	NA
	CCHFV	3	357	0.044	0.024	0.078	52.013	4.168	<0.001	0.269	0.102
	<i>Theileria</i> sp.	7	2381	0.033	0.007	0.139	98.126	327.182	<0.001	0.367	0.014
	Tick borne pathogens	13	4409	0.024	0.008	0.067	97.677	516/536	<0.001	0.076	0.008
Khorasan, South	<i>Anaplasma</i> sp.	2	159	0.299	0.129	0.552	88.666	8.823	0.115	NA	NA
	CCHFV	2	294	0.143	0.037	0.420	12.466	91.978	0.017	NA	NA
	Tick borne pathogens	5	553	0.176	0.089	0.317	87.984	33.288	<0.001	0.426	0.243
Khuzestan						NA					
Kohgiluyeh and Boyer-Ahmad						NA					
Kurdistan	<i>Borrelia</i> sp.	2	196	0.085	0.012	0.416	90.511	10.538	0.022	NA	NA
	Tick borne pathogens	5	6500	0.205	0.140	0.291	97.348	150.833	<0.001	0.462	0.240
Lorestan	<i>Theileria</i> sp.	5	583	0.125	0.064	0.228	83.837	24.748	<0.001	0.086	0.000
	Tick borne pathogens	11	1543	0.172	0.087	0.314	96.425	279.715	<0.001	0.061	0.064
Mazandaran	<i>Anaplasma</i> sp.	3	817	0.323	0.131	0.601	96.098	51.254	0.207	0.296	0.231
	CCHFV	2	188	0.047	0.023	0.090	15.618	1.185	<0.001	NA	NA
	<i>Theileria</i> sp.	2	40	0.210	0.015	0.824	85.552	6.921	0.009	NA	NA
	Tick borne pathogens	9	2157	0.100	0.031	0.277	97.765	357.961	0.001	0.754	0.049

Table 3. Continued.

Provinces	Pathogens	No. of studies	Sample size	Prevalence (pooled effect size)	95% CI		Heterogeneity		P values of prevalence	Publication bias	
					Lower	Upper	I <sup>2</sup> (%)	Q test		Begg's test (2 tailed P value)	Egger's test (2 tailed P value)
Qazvin	<i>Borrelia</i> sp. = Tick borne pathogens	4	943	0.077	0.020	0.253	94.702	56.624	0.001	0.734	0.993
Qom					NA						
Semnan	Tick borne pathogens	2	6031	0.041	0.036	0.046	0.000	0.010	<0.001	NA	NA
Sistan and Baluchestan	<i>Anaplasma</i> sp.	4	657	0.241	0.054	0.641	98.232	169.554	0.193	0.734	0.083
	<i>Coxiella</i> sp.	3	1988	0.041	0.015	0.103	89.324	18.734	<0.001	1.000	0.374
	CCHFV	3	289	0.040	0.018	0.090	28.217	2.786	<0.001	1.000	0.430
	<i>Theileria</i> sp.	3	549	0.122	0.097	0.152	29.684	2.844	<0.001	1.000	0.471
	Tick borne pathogens	14	3533	0.093	0.043	0.188	97.420	503.959	<0.001	1.000	0.392
Tehran	Tick borne pathogens	2	116	0.110	0.000	0.975	93.702	15.878	0.477	NA	NA
Yazd	<i>Theileria</i> sp.	2	500	0.015	0.001	0.299	82.719	5.787	0.014	NA	NA
	Tick borne pathogens	3	640	0.055	0.037	0.081	65.569	5.809	<0.001	0.296	0.142
Unspecified location	<i>Babesia</i> sp.	2	480	0.077	0.026	0.208	89.207	9.265	<0.001	NA	NA
	Tick borne pathogens	3	541	0.175	0.033	0.565	97.476	79.225	0.093	1.000	0.982
Iran (all collected data)	<i>Anaplasma</i> sp.	18	2373	0.235	0.151	0.347	96.596	498.733	<0.001	0.080	0.000
	<i>Babesia</i> sp.	17	6943	0.040	0.019	0.081	97.737	706.904	<0.001	0.010	0.000
	<i>Borrelia</i> sp.	15	5124	0.068	0.029	0.150	97.567	534.363	<0.001	1.000	0.289
	<i>Brucella</i> sp.				NA						
	CCHFV	31	4819	0.056	0.039	0.081	86.951	199.253	0.001	0.091	0.000
	<i>Coxiella</i> sp.	9	3753	0.065	0.030	0.138	96.738	245.246	<0.001	1.000	0.450
	<i>Ehrlichia</i> sp.	4	693	0.177	0.056	0.437	96.744	92.137	0.019	0.734	0.594
	<i>Rickettsia</i> sp.	2	125	0.283	0.029	0.839	96.370	27.552	0.480	NA	NA
	<i>Theileria</i> sp.	36	11076	0.093	0.067	0.129	96.157	910.777	<0.001	0.827	0.000
	Tick borne pathogens	135	35184	0.086	0.070	0.106	97.429	5211.303	<0.001	0.933	0.000

Note: In this analysis, each row of Table 1 was considered as an individual data. Furthermore, the sample size of each row of Table 1 was considered a separate sample size, and all pathogens were included. Provinces with less than two data were not included in meta-analysis. However, the pathogens detected in these provinces were calculated in Iran's total prevalence of pathogens section.

were Kurdistan (20.5%), Ardabil (18.6%), South Khorasan (17.6%), Lorestan (17.2%), East Azerbaijan (13.3%) and Fars (11.5%), respectively. Geographically, these provinces (except South Khorasan) are located in the western parts of Iran. Therefore, it can be concluded that although tick-borne pathogens have been reported from different regions of Iran, the western part of the country is more infected than other regions. This high prevalence can be justified due to high livestock population, common border with neighboring countries and traditional livestock holding methods with low hygiene.

In this analysis, 26 out of 31 provinces were surveyed regarding tick-borne pathogen detection in ticks; meanwhile, the status of infection of ticks to different pathogens remained unclear in five provinces: Alborz, Bushehr, Chaharmahal and Bakhtiari, Markazi, and Zanjan. Due to the importance of ticks and their impact on human and animal health, it is highly advisable to conduct studies concerning tick-borne diseases to clarify the status of these provinces. Vector surveillance seems to be vital for observing the

presence or occurrence of emerging and reemerging tick borne diseases in Iran and provides a preliminary warning for predicting probable epidemics.

In our analysis, *H. anatolicum* (20.35%), *R. sanguineus* (15.00%), and *R. bursa* (14.08%), were the most prevalent infected ticks in Iran. Genera of *Hyalomma* species have received much attention due to the role in the transmission of *Theileria* spp., *Babesia* spp., *Rickettsia* spp., and CCHFV. *R. sanguineus* (brown dog tick, kennel tick) is found worldwide with an interest toward warmer climates (tropics and sub-tropics)[121]. Dogs are specific host for *R. sanguineus*, however, it can be found on domestic ruminants and other vertebrates. Several pathogens such as *Ba. canis*, *Cx. burnetii*, *Eh. canis*, *Ri. conorii*, *Ri. rickettsii*, *Theileria* sp., *Anaplasma* sp., and CCHFV have been isolated from *R. sanguineus*[122-124]. *R. bursa* is common among livestock transmitting the protozoans *Ba. bigemina*, *Ba. caballi*, *Th. equi* and *Ba. bovis*[125]. Following these highly infected vectors, much lower prevalence levels were detected in *R. appendiculatus*, *H. schulzei*, *H. rufipes*, *H. aegyptium*



and *Boophilus* sp. These vectors should not be underestimated, as future investigations may reveal a high tendency of these species to transmit pathogens.

Controlling strategies against ticks and tick-borne diseases for prevention of significant losses due to both economic and public health problems are also seem to be important and helpful. Many attempts have been carried out for the control of ticks and tick-borne diseases[126]. Some other additional methods have been suggested: (1) livestock sheds should be checked regularly in terms of tick infestation; (2) different species of livestock should be held separately to avoid interspecies tick infection; (3) quarantine of newly purchased animals decreases the chance of tick transmission to existing animals; (4) periodic application of acaricide and chemotherapy according to regional and national guidelines is sometimes suggested; (5) clearance of vegetation cut off the connection between different stages of tick's life and disrupts their life cycle is also suggested; (6) some novel methods including application of vaccines against tick-borne pathogens, biological control, and genetically resistant livestock breeds are in the spotlight[127].

This investigation had some limitations: In the old classification of Iran provinces, some provinces are currently divided in two or more provinces, resulting in the less accuracy of the old literature, as they cover a larger area. In addition, access to the full text of some dissertations required a visit to the relevant center, which was very difficult due to the COVID-19 pandemic. In such cases, we missed some dissertations. Furthermore, the scientific name of some of tick species had changed since the publication of the associated papers, so we had to search with the old names as well.

In conclusion, the occurrence of at least 20 different pathogens (in 10 genera) in 28 species (in 9 genera) of ticks in 26 provinces of Iran, sheds light on the current status of the country in terms of tick-borne pathogens. Rate of infection to different pathogens in different regions, especially western parts of Iran, is a warning for public and animal health. Further investigations and persistent surveillance of vectors as well as vertebrate hosts will expand the chance of controlling tick-borne pathogens. In most parts of the meta-analysis concerning total pathogens of Iran, the results showed high heterogeneity ( $I^2 > 75\%$ ). Similarly, meta-analysis of separate provinces revealed high heterogeneity. This is not unexpected due to the variations associated with the different detection methods, sample size, geographical traits, location, time of the study, and population of interest. While the significance of a meta-analysis in regarding to the prevalence of tick-borne pathogens is undeniable, it is suggested that meta-analysis should not be an adequate alternative for large-scaled epidemiological studies due to heterogeneous approaches, regions and times of different studies.

## Conflict of interest statement

The authors declare that there is no conflict of interest.

## Authors' contributions

HB, MK, and ASJ planned for the study. HB, ASJ, MK, and MMS performed the literature search and data extraction. MK and ZT critically evaluated the manuscript. ASJ performed the meta-analysis. The final manuscript approved by all the authors.

## References

- [1] Salman MD, Tarrés-Call J. *Ticks and tick-borne diseases: Geographical distribution and control strategies in the Euro-Asia region*. CABI: Wallingford; 2013.
- [2] Brites-Neto J, Duarte KMR, Martins TF. Tick-borne infections in human and animal population worldwide. *Vet world* 2015; **8**(3): 301.
- [3] Nicholson WL, Sonenshine DE, Noden BH, Brown RN. Chapter 27-Ticks (Ixodidae). In: Mullen GR, Durden LA. *Medical and veterinary entomology*. 3rd ed. New York: Academic Press; 2019, p. 603-672.
- [4] de la Fuente J, Estrada-Pena A, Venzal JM, Kocan KM, Sonenshine DE. Overview: Ticks as vectors of pathogens that cause disease in humans and animals. *Front Biosci* 2008; **13**(13): 6938-6946.
- [5] Abubakar M, Perera PK, Iqbal A, Manzoor S. Introductory chapter: Ticks and tick-borne pathogens. In: *Ticks and tick-borne pathogens*. London: IntechOpen; 2018.
- [6] Dantas-Torres F, Chomel BB, Otranto D. Ticks and tick-borne diseases: A One Health perspective. *Trends Parasitol* 2012; **28**(10): 437-446.
- [7] Mashebe P, Lyaku JR, Mause F. Occurrence of ticks and tick-borne diseases of livestock in Zambezi region: A review. *J Agric Sci* 2014; **6**(2): 142.
- [8] Hurtado OJB, Giraldo-Ríos C. Economic and health impact of the ticks in production animals. In: *Ticks and tick-borne pathogens*. London: IntechOpen; 2018.
- [9] Rochlin I, Toledo A. Emerging tick-borne pathogens of public health importance: A mini-review. *J Med Microbiol* 2020; **69**(6): 781.
- [10] Estrada-Peña A, Ayllón N, De La Fuente J. Impact of climate trends on tick-borne pathogen transmission. *Front Physiol* 2012; **3**: 64.
- [11] Kiyani Haftlang K. *The book of Iran: A survey of the geography of Iran*. Tehran: Alhoda UK; 2003.
- [12] Randolph SE. Tick ecology: Processes and patterns behind the epidemiological risk posed by ixodid ticks as vectors. *Parasitology* 2004; **129**(S1): S37.
- [13] Hosseini-Chegeni A, Tavakoli M, Telmadarraiy Z. The updated list of ticks (Acari: Ixodidae & Argasidae) occurring in Iran with a key to the

- identification of species. *Syst Appl Acarol* 2019; **24**(11): 2133-2166.
- [14]Gholamreza S, Somaieh M, Roya S, Alireza B, Ghazale A, Yasin B. First detection of *Babesia ovis* in *Dermacentor* spp. in Ardabil area, northwest of Iran. *J Vector Borne Dis* 2017; **54**(3): 277.
- [15]Arjmand Yamchi J, Tavassoli M. Survey on infection rate, vectors and molecular identification of *Theileria annulata* in cattle from North West, Iran. *J Parasit Dis* 2016; **40**(3): 1071-1076.
- [16]Aghaei A, Ghazinezhad B, Naddaf SR. Detection of *Borrelia* DNA in *Ornithodoros tholozani* ticks and their eggs. *J Med Microbiol Infect Dis* 2014; **2**(3): 118-120.
- [17]Arshi SH, Majidpour A, Sadeghi H, Emdadi D, Asmar M, Derakhshan MH. Relapsing fever in Ardabil, a northwestern province of Iran. *Arch Iran Med* 2002; **5**(3): 141-145.
- [18]Telmadarraiy Z, Ghiasi SM, Moradi M, Vatandoost H, Eshraghian MR, Faghihi F, et al. A survey of Crimean-Congo haemorrhagic fever in livestock and ticks in Ardabil Province, Iran during 2004-2005. *Scand J Infect Dis* 2010; **42**(2): 137-141.
- [19]Esmailnejad B, Gharekhani J, Rezaei ASH. Molecular detection of *Coxiella burnetii* in ticks isolated from goats of Meshkin-Shahr County, Ardabil Province, Iran. *Nov Biol Reper* 2020; **7**(3): 315-321.
- [20]Khazeni A, Telmadarraiy Z, Oshaghi MA, Mohebbali M, Zarei Z, Abtahi SM. Molecular detection of *Ehrlichia canis* in ticks population collected on dogs in Meshkin-Shahr, Ardebil Province, Iran. *J Biomed Sci Eng* 2013; **6**: 1-5.
- [21]Abdoli R, Bakhshi H, Kheirandish S, Faghihi F, Hosseini-Chegeni A, Oshaghi MA, et al. Circulation of Brucellaceae, *Anaplasma* and *Ehrlichia* spp. in borderline of Iran, Azerbaijan, and Armenia. *Asian Pac J Trop Med* 2021; **14**: 223-230.
- [22]Tajedin L, Bakhshi H, Faghihi F, Telmadarraiy Z. High infection of *Anaplasma* and *Ehrlichia* spp. among tick species collected from different geographical locations of Iran. *Asian Pacific J Trop Dis* 2016; **6**(10): 787-792.
- [23]Nadim A, Khanjani M, Hosseini-Chegeni A, Telmadarraiy Z. Identity and microbial agents related to *Dermacentor marginatus* Sulzer (Acari: Ixodidae) with a new record of *Rickettsia slovaca* (Rickettsiales: Rickettsiaceae) in Iran. *Syst Appl Acarol* 2021; **26**(2): 367-378.
- [24]Jafarbekloo A, Ramzgouyan MR, Shirian S, Tajedin L, Bakhshi H, Faghihi F, et al. Molecular characterization and phylogenetic analysis of *Theileria* spp. and *Babesia* spp. isolated from various ticks in southeastern and northwestern regions of Iran. *Vector-Borne Zoonotic Dis* 2018; **18**(11): 595-600.
- [25]Shafei E, Dayer MS, Telmadarraiy Z. Molecular epidemiology of Crimean-Congo hemorrhagic fever virus in ticks in northwest of Iran. *J Entomol Zool Stud* 2016; **4**(5): 150-154.
- [26]Rajabi S, Esmailnejad B, Tavassoli M. A molecular study on *Babesia* spp. in cattle and ticks in West-Azerbaijan province, Iran. *Vet Res Forum* 2017; **8**(4): 299-306.
- [27]Esmailnejad B, Tavassoli M, Asri-Rezaei S, Dalir-Naghadeh B, Mardani K, Jalilzadeh-Amin G, et al. PCR-based detection of *Babesia ovis* in *Rhipicephalus bursa* and small ruminants. *J Parasitol Res* 2014; **2014**: 1-6. doi: 10.1155/2014/294704.
- [28]Tavassoli M, Tabatabaei M, Mohammadi M, Esmailnejad B, Mohamadpour H. PCR-based detection of *Babesia* spp. infection in collected ticks from cattle in west and north-west of Iran. *J Arthropod Borne Dis* 2013; **7**(2): 132.
- [29]Mohammadi SM, Esmailnejad B, Jalilzadeh-Amin G. Molecular detection, infection rate and vectors of *Theileria lestoquardi* in goats from West Azerbaijan province, Iran. *Vet Res Forum* 2017; **8**(2): 139-144.
- [30]Abdigoudarzi M. Detection of naturally infected vector ticks (Acari: Ixodidae) by different species of *Babesia* and *Theileria* agents from three different enzootic parts of Iran. *J Arthropod Borne Dis* 2013; **7**(2): 164-172.
- [31]Spitalska E, Namavari MM, Hosseini MH, Shad-Del F, Amrabadi OR, Sparagano OAE. Molecular surveillance of tick-borne diseases in Iranian small ruminants. *Small Rumin Res* 2005; **57**(2-3): 245-248.
- [32]Farhadpour F, Telmadarraiy Z, Chinikar S, Akbarzadeh K, Moemenbellah-Fard MD, Faghihi F, et al. Molecular detection of Crimean-Congo haemorrhagic fever virus in ticks collected from infested livestock populations in a New Endemic Area, South of Iran. *Trop Med Int Heal* 2016; **21**(3): 340-347.
- [33]Yaghfoori S, Razmi G, Heidarpour M. Molecular detection of *Theileria* spp. in sheep and vector ticks in Fasa and Kazeroun areas, Fars Province, Iran. *Arch Razi Institute* 2013; **68**(2): 159-164.
- [34]Hosseini-Chegeni A, Tavakoli M, Goudarzi GH, Telmadarraiy Z, Sharifdini M, Faghihi F, et al. Molecular detection of *Anaplasma marginale* and *Anaplasma ovis* (Rickettsiales: Anaplasmataceae) in ixodid tick species in Iran. *Arch Razi Inst* 2020; **75**(1): 39-46.
- [35]Naddaf SR, Mahmoudi A, Ghasemi A, Rohani M, Mohammadi A, Ziapour SP, et al. Infection of hard ticks in the Caspian Sea littoral of Iran with Lyme borreliosis and relapsing fever borreliae. *Ticks Tick Borne Dis* 2020; **11**(6): 101500.
- [36]Hosseini-Chegeni A, Tavakoli M, Telmadarraiy Z, Sedaghat MM, Faghihi F. Detection of a *Brucella*-like (Alphaproteobacteria) bacterium in *Boophilus* spp. (Acari: Ixodidae) from Iran. *J Med Microbiol Infect Dis* 2017; **5**(3): 66-68.
- [37]Bekloo AJ, Bakhshi H, Soufizadeh A, Sedaghat MM, Bekloo RJ, Ramzgouyan MR, et al. Ticks circulate *Anaplasma*, *Ehrlichia*, *Babesia* and *Theileria* parasites in North of Iran. *Vet Parasitol* 2017; **248**: 21-24.
- [38]Sedaghat MM, Sarani M, Chinikar S, Telmadarraiy Z, Moghaddam AS, Azam K, et al. Vector prevalence and detection of Crimean-Congo haemorrhagic fever virus in Golestan Province, Iran. *J Vector Borne Dis* 2017; **54**(4): 353.
- [39]Vatandoost H, Ghaderi A, Javadian E, Nia AHZ, Rassi Y, Piazak N, et al. Distribution of soft ticks and their infection with *Borrelia* in Hamadan province, Iran. *Iran J Public Health* 2003; **32**(1): 22-24.
- [40]Shahraki G, Asmar M. Study on distribution of Arasid-ticks and their infection to *Borrelia persica* in indoor resting places of Hamadan. [Online]. Available from: <http://sjh.umsha.ac.ir/article-1-1065-en.html>. [Accessed on 8 November 2021].
- [41]Taher M, Dayer M, Jalali T, Khakifirouz S, Telmadarraiy Z, Salehi-Vaziri M. Molecular epidemiology of Crimean-Congo hemorrhagic fever virus in ticks collected from western Iran. *Asian Biomed* 2016; **10**(6): 603-607.



- [42]Tahmasebi F, Ghiasi SM, Mostafavi E, Moradi M, Piazak N, Mozafari A, et al. Molecular epidemiology of Crimean-Congo hemorrhagic fever virus genome isolated from ticks of Hamadan province of Iran. *J Vector Borne Dis* 2010; **47**(4): 211-216.
- [43]Telmadarraiy Z, Moradi AR, Vatandoost H, Mostafavi E, Oshaghi MA, Zahirnia AH, et al. Crimean-Congo hemorrhagic fever: A seroepidemiological and molecular survey in Bahar, Hamadan province of Iran. *Asian J Anim Vet Adv* 2008; **3**(5): 321-327.
- [44]Sharifinia N, Rafinejad J, Hanafi-Bojd AA, Chinikar S, Piazak N, Baniardalani M, et al. Hard ticks (Ixodidae) and Crimean-Congo hemorrhagic fever virus in south west of Iran. *Acta Med Iran* 2015; **53**(3): 177-181.
- [45]Biglari P, Chinikar S, Belqeisizadeh H, Telmadarraiy Z, Mostafavi E, Ghaffari M, et al. Phylogeny of tick-derived Crimean-Congo hemorrhagic fever virus strains in Iran. *Ticks Tick Borne Dis* 2016; **7**(6): 1216-1221.
- [46]Ranjbar R, Anjomruz M, Enayati AA, Khoobdel M, Rafinejad A, Rafinejad J. *Anaplasma* infection in ticks in southeastern region of Iran. *J Arthropod Borne Dis* 2020; **14**(2): 126-133.
- [47]Akhtardanesh B, Saberi M, Nurollahifard SR, Aghazamani M. Molecular detection of *Babesia* spp. in tick-infested dogs in Southeastern Iran. *J Dis Glob Heal* 2016; **8**(2): 72-77.
- [48]Salehi-Vaziri M, Vatandoost H, Sanei-Dehkordi A, Fazlalipour M, Pouriayevali MH, Jalali T, et al. Molecular assay on detection of Crimean Congo hemorrhagic fever (CCHF) virus in ixodid ticks collected from livestock in slaughterhouse from South of Iran. *J Arthropod Borne Dis* 2020; **14**(3): 286-292.
- [49]Khakifirouz S, Mowla SJ, Baniasadi V, Fazlalipour M, Jalali T, Mirghiasi SM, et al. No detection of Crimean Congo hemorrhagic fever (CCHF) virus in ticks from Kerman Province of Iran. *J Med Microbiol Infect Dis* 2018; **6**(4): 108-111.
- [50]Khalili M, Rezaei M, Akhtardanesh B, Abiri Z, Shahheidaripour S. Detection of *Coxiella burnetii* (Gammaproteobacteria: Coxiellaceae) in ticks collected from infested dogs in Kerman, Southeast of Iran. *Persian J Acarol* 2018; **7**(1): 93-110. doi: 10.22073/pja.v7i1.30699.
- [51]Fard SN, Khalili M. PCR-detection of *Coxiella burnetii* in ticks collected from sheep and goats in Southeast Iran. *Iran J Arthropod Borne Dis* 2011; **5**(1): 1-6.
- [52]Motaghpisheh S, Akhtardanesh B, Ghanbarpour R, Aflatoonian MR, Khalili M, Nourollahifard SR, et al. *Ehrlichiosis* in household dogs and parasitized ticks in Kerman-Iran: Preliminary zoonotic risk assessment. *J Arthropod Borne Dis* 2016; **10**(2): 245-251.
- [53]Mohammadian M, Chinikar S, Telmadarraiy Z, Vatandoost H, Oshaghi MA, Hanafi-Bojd AA, et al. Molecular assay on Crimean Congo hemorrhagic fever virus in ticks (Ixodidae) collected from Kermanshah Province, Western Iran. *J Arthropod Borne Dis* 2016; **10**(3): 381-391.
- [54]Rahmani-Varmale M, Tavassoli M, Esmailnejad B. Molecular detection and differentiation of *Theileria lestoquardi*, *Th. ovis* and *Th. annulata* in blood of goats and ticks in Kermanshah Province, Iran. *J Arthropod Borne Dis* 2019; **13**(3): 297-309.
- [55]Abedi V, Razmi G, Seifi H, Naghibi A. Molecular and serological detection of *Theileria equi* and *Babesia caballi* infection in horses and ixodid ticks in Iran. *Ticks Tick Borne Dis* 2014; **5**(3): 239-244.
- [56]Seidabadi M, Razmi G, Naghibi A. Molecular detection of *Babesia* spp. in sheep and vector ticks in North Khorasan province, Iran. *Iran J Vet Med* 2014; **8**(1): 35-39.
- [57]Saghafipour A, Mousazadeh-Mojarrad A, Arzamani N, Telmadarraiy Z, Rajabzadeh R, Arzamani K. Molecular and seroepidemiological survey on Crimean-Congo hemorrhagic fever virus in Northeast of Iran. *Med J Islam Repub Iran* 2019; **33**: 41. doi: 10.34171/mjiri.33.41.
- [58]Champour M, Chinikar S, Mohammadi G, Razmi G, Mostafavi E, Shah-Hosseini N, et al. Crimean-Congo hemorrhagic fever in the one-humped camel (*Camelus dromedarius*) in East and Northeast of Iran. *J Arthropod Borne Dis* 2016; **10**(2): 168-177.
- [59]Rashidi A, Razmi G. Molecular detection of *Theileria* spp. in sheep and vector ticks in the North Khorasan Province, Iran. *Trop Anim Health Prod* 2012; **45**(1): 299-303.
- [60]Khodaverdi Azghandi M, Razmi G. Identification of *Babesia* and *Theileria* species in goats and ticks with smear observation and molecular examination in Mashhad, Khorasan Razavi province, Iran. *J Vet Res* 2015; **70**(1): 1-5.
- [61]Razmi G, Pourhosseini M, Yaghfoury S, Rashidi A, Seidabadi M. Molecular detection of *Theileria* spp. and *Babesia* spp. in sheep and ixodid ticks from the northeast of Iran. *J Parasitol* 2013; **99**(1): 77-81.
- [62]Shayeghi M, Piazak N, Gollampour A, Nasirian H, Abolhassani M. Tick-borne relapsing fever in Sabzevar (Khorasan Razavi Province), North-Eastern Iran. *Bangladesh J Med Sci* 2016; **15**(4): 551-555.
- [63]Maghsood H, Nabian S, Shayan P, Jalali T, Darbandi MS, Ranjbar MM. Molecular epidemiology and phylogeny of Crimean-Congo haemorrhagic fever (CCHF) virus of ixodid ticks in Khorasan Razavi Province of Iran. *J Arthropod Borne Dis* 2021; **14**(4): 400-407.
- [64]Fakoorziba MR, Naddaf-Sani AA, Moemenbellah-Fard MD, Azizi K, Ahmadnia S, Chinikar S. First phylogenetic analysis of a Crimean-Congo hemorrhagic fever virus genome in naturally infected *Rhipicephalus appendiculatus* ticks (Acari: Ixodidae). *Arch Virol* 2015; **160**(5): 1197-1209.
- [65]Razmi GR, Ebrahimzadeh E, Aslani MR. A study about tick vectors of bovine theileriosis in an endemic region of Iran. *J Vet Med Ser B* 2003; **50**(6): 309-310.
- [66]Razmi G, Yaghfoori S. Molecular surveillance of *Theileria ovis*, *Theileria lestoquardi* and *Theileria annulata* infection in sheep and ixodid ticks in Iran. *Onderstepoort J Vet Res* 2013; **80**(1): 635. doi: 10.4102/ojvr.v80i1.635.
- [67]Jafari A. *Epidemiology and molecular detection of Crimean-Congo hemorrhagic fever (CCHF), Coxiella burnetii and Anaplasma spp. in hard ticks (Ixodidae) in the South Khorasan regions*. DVM. Thesis. Zabol, Iran: University of Zabol; 2020.
- [68]Asadollahi Z, Jalali MHR, Alborzi A, Hamidinejat H. Detection of *Theileria*-like organisms in *Hyalomma* ticks (Acarina: Ixodidae) in Khuzestan, Iran. *Sci Parasitol* 2018; **19**(1-2): 34-39.
- [69]Hosseini Z, Salehi Vaziri M, Ahmadnia S, Fakoorziba MR, Jalali T, Telmadarraiy Z, et al. Hard ticks infesting domestic ruminants, species composition and infection with Crimean-Congo hemorrhagic fever virus

- in a highland province, SW Iran. *J Heal Sci Surveill Syst* 2019; **7**(2): 52-59.
- [70] Hasheminasab SS, Moradi P, Wright I. A four year epidemiological and chemotherapy survey of babesiosis and theileriosis, and tick vectors in sheep, cattle and goats in Dehgolan, Iran. *Ann Parasitol* 2018; **64**(1): 43-48. doi:10.17420/ap6401.131.
- [71] Moemenbellah-Fard MD, Benafshi O, Rafinejad J, Ashraf H. Tick-borne relapsing fever in a new highland endemic focus of western Iran. *Ann Trop Med Parasitol* 2009; **103**(6): 529-537.
- [72] Banafshi O, Rafinejad J, Esmaeinasab N. Study of the spread of soft ticks (Argasidae) in indoor areas and the study of infection of *Ornithodoros tolozani* with *Borrelia persica* in Bijar city of Kurdistan province. *Sci J Kurdistan Univ Med Sci* 2004; **8**(31): 50-58.
- [73] Fakoorziba MR, Golmohammadi P, Moradzadeh R, Moemenbellah-Fard MD, Azizi K, Davari B, et al. Reverse transcription PCR-based detection of Crimean-Congo hemorrhagic fever virus isolated from ticks of domestic ruminants in Kurdistan Province of Iran. *Vector Borne Zoonotic Dis* 2012; **12**(9): 794-799.
- [74] Chegeni AH, Tavakoli M. *Aegyptianella pullorum* (Rickettsiales: Anaplasmataceae) in tick *Argas persicus* (Acari: Argasidae) from Iran: A preliminary assessment. *Persian J Acarol* 2018; **7**(3): 307-311. doi:10.22073/pja.v7i3.37407.
- [75] Chegeni AH, Telmadarraiy Z, Tavakoli M, Faghihi F. Molecular detection of *Borrelia anserina* in *Argas persicus* (Acari: Argasidae) ticks collected from Lorestan province, west of Iran. *Persian J Acarol* 2017; **6**(4): 287-297. doi:10.22073/pja.v6i4.28372.
- [76] Hosseini-Chegeni A, Kayedi MH. Molecular detection of *Coxiella* (Gammaproteobacteria: Coxiellaceae) in *Argas persicus* and *Alveonatus canestrinii* (Acari: Argasidae) from Iran. *Microb Pathog* 2020; **139**: 103902.
- [77] Kayedi MH, Chinikar S, Mostafavi E, Khakifirouz S, Jalali T, Hosseini-Chegeni A, et al. Crimean-Congo hemorrhagic fever virus clade IV (Asia 1) in ticks of Western Iran. *J Med Entomol* 2015; **52**(5): 1144-1149.
- [78] Kooshki H, Goudarzi G, Faghihi F, Telmadarraiy Z, Edalat H, Hosseini-Chegeni A. The first record of *Rickettsia hoogstraalii* (Rickettsiales: Rickettsiaceae) from *Argas persicus* (Acari: Argasidae) in Iran. *Syst Appl Acarol* 2020; **25**(9): 1611-1617.
- [79] Nasser H-R, Saeed H, Mohammad A. Molecular detection of *Theileria ovis* and *Th. lestoquardi* in vector ticks in Lorestan province, Iran. *Int J Biosci* 2014; **4**(12): 78-83.
- [80] Hashemi S, Estaki Oregani K. Molecular identification of *Theileria ovis* and *Th. lestoquardi* in vector ticks of Ixodidae family in Lorestan province. *Iran Vet J* 2015; **11**(3): 97-104.
- [81] Pazhoom F, Ebrahimzade E, Shayan P, Nabian S. *Anaplasma* spp. identification in hard ticks of Iran: First report of *Anaplasma bovis* in *Haemaphysalis inermis*. *Acarologia* 2016; **56**(4): 497-504.
- [82] Hosseini-Vasoukolaei N, Oshaghi MA, Shayan P, Vatandoost H, Babamahmoudi F, Yaghoobi-Ershadi MR, et al. *Anaplasma* infection in ticks, livestock and human in Ghaemshahr, Mazandaran Province, Iran. *J Arthropod Borne Dis* 2014; **8**(2): 204-211.
- [83] Bashiribod H. First molecular detection of *Anaplasma phagocytophilum* in *Ixodes ricinus* ticks in Iran. *J Med Sci* 2004; **4**(4): 282-286.
- [84] Bashiribod H, Rahbarian N, Eslami G, Kazemi B, Jannatsharif E, Mahmoudirad M, et al. Prevalence of *Coxiella burnetii* in human, animal hosts and hard ticks in West Mazandaran Province Iran, 2003-2004. *Pajouhesh Dar Pezeshki* 2008; **32**(3): 253-257.
- [85] Hosseini-Vasoukolaei N, Chinikar S, Telmadarraiy Z, Faghihi F, Hosseini-Vasoukolaei M. Serological and molecular epidemiology of Crimean-Congo hemorrhagic fever in Ghaemshahr county in Mazandaran province, Iran. *Trop Biomed* 2016; **33**(4): 807-813.
- [86] Zakkyeh T, Mohammad Ali O, Nasibeh HV, Mohammad Reza YE, Farhang B, Fatemeh M. First molecular detection of *Theileria ovis* in *Rhipicephalus sanguineus* tick in Iran. *Asian Pac J Trop Med* 2012; **5**(1): 29-32. doi: 10.1016/S1995-7645(11)60240-X.
- [87] Aghighi Z, Assmar M, Piazak N, Javadian E, Seyedi RMA, Kia EB, et al. Distribution of soft ticks and their natural infection with *Borrelia* in a focus of relapsing fever in Iran. *J Arthropod Borne Dis* 2007; **1**(2): 14-18.
- [88] Barmaki A, Rafinejad J, Vatandoost H, Telmadarraiy Z, Mohtarami F, Leghaei SH, et al. Study on presence of *Borrelia persica* in soft ticks in Western Iran. *Iran J Arthropod Borne Dis* 2010; **4**(2): 19-25.
- [89] Telmadarraiy Z, Saghafipour A, Farzinnia B, Chinikar S. Molecular detection of Crimean-Congo hemorrhagic fever virus in ticks in Qom Province, Iran, 2011-2012. *Iran J Virol* 2012; **6**(3): 13-18.
- [90] Nekooyi H, Asmar M, Amirkhani A, Piyazak N. Geographical distribution of ticks in Semnan province and the rate of infection of soft ticks with *Borrelia*. *Iran J Heal* 1999; **4**(1): 103-110.
- [91] Faghihi F, Telmadarraiy Z, Chinikar S, Nowotny N, Fooks AR, Shahhosseini N. Spatial and phylodynamic survey on Crimean-Congo hemorrhagic fever virus strains in northeast of Iran. *Jundishapur J Microbiol* 2018; **11**(3): e59412. doi: 10.5812/ijm.59412.
- [92] Choubdar N, Karimian F, Koosha M, Nejati J, Oshaghi MA. *Hyalomma* spp. ticks and associated *Anaplasma* spp. and *Ehrlichia* spp. on the Iran-Pakistan border. *Parasit Vectors* 2021; **14**: 469.
- [93] Khodadadi N, Nabavi R, Sarani A, Saadati D, Ganjali M, Mihalca AD, et al. Identification of *Anaplasma marginale* in long-eared hedgehogs (*Hemiechinus auritus*) and their *Rhipicephalus turanicus* ticks in Iran. *Ticks Tick Borne Dis* 2021; **12**(2): 101641.
- [94] Asadollahi S. *Epidemiology & molecular detection of Crimean-Congo hemorrhagic fever (CCHF), Coxiella burnetii and Anaplasma spp. in hard ticks (Ixodidae) in the Sistan regions*. DVM. Thesis. Zabol, Iran: University of Zabol; 2020.
- [95] Jafarbekloo A, Bakhshi H, Faghihi F, Telmadarraiy Z, Khazeni A, Oshaghi MA, et al. Molecular detection of *Anaplasma* and *Ehrlichia* infection in ticks in borderline of Iran-Afghanistan. *J Biomed Sci Eng* 2014; **7**(11): 919-926. doi: 10.4236/jbise.2014.711089.
- [96] Ghashghaei O, Fard SRN, Khalili M, Sharifi H. A survey of ixodid ticks feeding on cattle and molecular detection of *Coxiella burnetii* from ticks in Southeast Iran. *Turkish J Vet Anim Sci* 2017; **41**(1): 46-50.
- [97] Fard SRN, Ghashghaei OO, Khalili M, Sharifi H. Tick diversity and detection of *Coxiella burnetii* in tick of small ruminants using nested Trans PCR in southeast Iran. *Trop Biomed* 2016; **33**(3): 506-511.
- [98] Hormozzayi H. *Molecular study of Ehrlichia infection in Rhipicephalus*

- sanguineus* ticks isolated from dogs in Zabol city. DVM. Thesis. Kerman, Iran: University of Kerman; 2017.
- [99]Shahhosseini N, Jafarbekloo A, Telmadarraiy Z, Chinikar S, Haeri A, Nowotny N, et al. Co-circulation of Crimean-Congo hemorrhagic fever virus strains Asia 1 and 2 between the border of Iran and Pakistan. *Heliyon* 2017; **3**(11): e00439.
- [100]Mehrvan A, Moradi M, Telmadarraiy Z, Mostafavi E, Moradi AR, Khakifirouz S, et al. Molecular detection of Crimean-Congo haemorrhagic fever (CCHF) virus in ticks from southeastern Iran. *Ticks Tick Borne Dis* 2013; **4**(1-2): 35-38.
- [101]Zarei F, Ganjali M, Nabavi R. Identification of *Theileria* species in sheep and vector ticks using PCR method in Zabol, Eastern Iran. *J Arthropod Borne Dis* 2019; **13**(1): 76-82.
- [102]Razmi GR, Hosseini M, Aslani MR. Identification of tick vectors of ovine theileriosis in an endemic region of Iran. *Vet Parasitol* 2003; **116**(1): 1-6.
- [103]Talaie P, Sedaghat MM, Mostafavi E, Telmadarraiy Z, Rouhani M, Salehi-Vaziri M. A Survey of Crimean-Congo hemorrhagic fever virus in ticks of Shahr-e Ray, Iran, 2016-2017. *J Med Microbiol Infect Dis* 2020; **8**(2): 56-59.
- [104]Habibi G, Imani A, Afshari A, Bozorgi S. Detection and molecular characterization of *Babesia canis vogeli* and *Theileria annulata* in free-ranging dogs and ticks from Shahriar County, Tehran Province, Iran. *Iran J Parasitol* 2020; **15**(3): 321-331. doi: 10.18502/ijpa.v15i3.4196.
- [105]Yaser SA, Sadegh C, Zakkyeh T, Hassan V, Maryam M, Ali OM, et al. Crimean-Congo hemorrhagic fever: A molecular survey on hard ticks (Ixodidae) in Yazd Province, Iran. *Asian Pac J Trop Med* 2011; **4**(1): 61-63.
- [106]Khodabandeh S, Razmi G. Molecular detection of *Theileria* species and its vectors in cattle of Yazd by Semi-nested PCR method. *J Vet Res* 2015; **70**(3): 249-253.
- [107]Shayan P, Hooshmand E, Rahbari S, Nabian S. Determination of *Rhipicephalus* spp. as vectors for *Babesia ovis* in Iran. *Parasitol Res* 2007; **101**(4): 1029-1033.
- [108]Hosseini-Chegeni A, Telmadarraiy Z, Faghihi F. Molecular detection of spotted fever group *Rickettsia* (Rickettsiales: Rickettsiaceae) in ticks of Iran. *Razi Vaccine Serum Res Inst* 2019; **75**(3): 317-325.
- [109]Ybañez AP, Inokuma H. *Anaplasma* species of veterinary importance in Japan. *Vet World* 2016; **9**(11): 1190-1196. doi: 10.14202/vetworld.2016.1190-1196.
- [110]Yang J, Liu Z, Niu Q, Liu J, Han R, Liu G, et al. Molecular survey and characterization of a novel *Anaplasma* species closely related to *Anaplasma capra* in ticks, northwestern China. *Parasit Vectors* 2016; **9**(1): 1-5.
- [111]Rymaszevska A, Grenda S. Bacteria of the genus *Anaplasma*-characteristics of *Anaplasma* and their vectors: A review. *Vet Med* 2008; **53**(11): 573-584.
- [112]Hanafi-Bojd AA, Jafari S, Telmadarraiy Z, Abbasi-Ghahramanloo A, Moradi-Asl E. Spatial distribution of ticks (Arachniada: Argasidae and Ixodidae) and their infection rate to Crimean-Congo hemorrhagic fever Virus in Iran. *J Arthropod Borne Dis* 2021; **15**(1): 41-59.
- [113]Salehi-Vaziri M, Baniasadi V, Jalali T, Mirghiasi SM, Azad-Manjiri S, Zarandi R, et al. The first fatal case of Crimean-Congo hemorrhagic fever caused by the AP92-like strain of the Crimean-Congo hemorrhagic fever virus. *Jpn J Infect Dis* 2016; **69**(4): 344-346.
- [114]Mostafavi E, Haghdoost A, Khakifirouz S, Chinikar S. Spatial analysis of Crimean Congo hemorrhagic fever in Iran. *Am J Trop Med Hyg* 2013; **89**(6): 1135-1141.
- [115]Cutler SJ, Ruzic-Sabljić E, Potkonjak A. Emerging borreliae-expanding beyond Lyme borreliosis. *Mol Cell Probes* 2017; **31**: 22-27.
- [116]Asl HM, Goya MM, Vatandoost H, Zahraei SM, Mafi M, Asmar M, et al. The epidemiology of tick-borne relapsing fever in Iran during 1997-2006. *Travel Med Infect Dis* 2009; **7**(3): 160-164.
- [117]Rezaei A, Gharibi D, Pourmahdi Borujeni M, Mosallanejad B. Seroprevalence of Lyme disease and Q fever in referred dogs to veterinary hospital of Ahvaz. *Iran Vet J* 2016; **11**(4): 34-41.
- [118]Esmaili S, Golzar F, Ayubi E, Naghili B, Mostafavi E. Acute Q fever in febrile patients in northwestern of Iran. *PLoS Negl Trop Dis* 2017; **11**(4): e0005535.
- [119]Mobarez AM, Amiri FB, Esmaili S. Seroprevalence of Q fever among human and animal in Iran; A systematic review and meta-analysis. *PLoS Negl Trop Dis* 2017; **11**(4): e0005521.
- [120]Rezaei M, Khalili M, Akhtardanesh B, Shahheidaripour S. Q fever in dogs: An emerging infectious disease in Iran. *J Med Bacteriol* 2016; **5**(1-2): 1-6.
- [121]Kumar B, Manjunathachar HV, Ghosh S. A review on *Hyalomma* species infestations on human and animals and progress on management strategies. *Heliyon* 2020; **6**(12): e05675.
- [122]René-Martellet M, Minard G, Massot R, Moro CV, Chabanne L, Mavingui P. Bacterial microbiota associated with *Rhipicephalus sanguineus* (sl) ticks from France, Senegal and Arizona. *Parasit Vectors* 2017; **10**(1): 1-10.
- [123]Dantas-Torres F. The brown dog tick, *Rhipicephalus sanguineus* (Latreille, 1806)(Acari: Ixodidae): From taxonomy to control. *Vet Parasitol* 2008; **152**(3-4): 173-185.
- [124]Dantas-Torres F. Biology and ecology of the brown dog tick, *Rhipicephalus sanguineus*. *Parasit Vectors* 2010; **3**(1): 26. doi: 10.1186/1756-3305-3-26.
- [125]Vatanserver Z. *Rhipicephalus bursa* Canestrini and Fanzago, 1878 (Figs. 117-119). In: *Ticks of Europe and North Africa*. Cham: Springer International Publishing; 2017, p. 299-303.
- [126]Vatandoost H, Moradi Asl E, Telmadarraiy Z, Mohebbali M, Masoumi Asl H, Abai MR, et al. Field efficacy of flumethrin pour-on against livestock ticks in Iran. *Int J Acarol* 2012; **38**(6): 457-464.
- [127]Muhammad G, Naureen A, Firyal S, Saqib M. Tick control strategies in dairy production medicine. *Pak Vet J* 2008; **28**(1): 43-50.