Mini Review Article

Therapeutic Potential of Glucose Regulated Protein 78 in Cancer

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Abstract

Endoplasmic reticulum has a critical role in the synthesis and folding of secretory and membrane proteins. High accumulation of proteins in ER activates the unfolded protein response and glucose regulated protein 78 or GPR78 plays an essential role in this pathway. Unfolded protein response is activated in cancerous cells due to their adverse condition to survive and it has been shown that GRP78 can be expressed in tumor cell membrane. Overexpression and localization of GRP78 makes it a suitable target for the treatment of cancer. This review describes cellular localization, biological function, and role of GRP78 in cancer induction. Methods for tumor inhibition via GRP78 are also discussed.

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Introduction

The endoplasmic reticulum (ER) has an essential role in the synthesis and folding of secretory and membrane proteins. Due to accumulation of unfold proteins over the capacity of the ER, the unfolded protein response (UPR), as a protective process is activated leading to translational attenuation, up-regulation of chaperones and folding enzymes and enhanced ER-associated degradation of misfolded proteins. Several mediators such as PKR-like

ER-associated kinase (PERK), IRE1/X-box binding protein-1 (XBP-1), and activating transcription factor-6 (ATF6) are inVolved in this process [1-4]. Severity of ER stresss pecifies the UPR which can cause cell death by activating the ER mediated apoptotic pathways, as well as coupling to the mitochondrial pathways [4, 5]. ER stress can also induce autophagy, a cellular degradation process implicated in both cell death and survival [6].

GRP78 is a 78 kDa molecular chaperone with the binding ability to hydrophobic patches on nascent polypeptides in the ER to prevent their aggregation ,thereby promoting folding of entire polypeptides in to proper native con formations [7]. Due to this function , GRP78 is expressed in all mammalian cells as one of the initial components in the signaling cascade of the UPR to inhibit aggregation of unfolded proteins [7].

The available pool of GRP78 in the ER induces secondary signaling mediators that directly act to prevent the accumulation of unfolded proteins in the cell or across the nuclear membrane [8]. GRP78 shows response to Ca^{2+} , glucose and energy depletion, accumulation of unfolded proteins in the ER. These events are regulated by GRP78 interactions with some of the secondary signaling proteins such as inositol requiring kinase 1 (IRE1), PKR-like ER-

associated kinase (PERK), and activating transcription factor 6 (ATF6) [2, 9].

Localization of GRP78 in the cell

The presence of the KDEL retention motif on C-terminus of GRP78 makes this protein as an ER lumen-localized chaperone [10]. However, that the presence of GRP78 in nucleus and mitochondria has also been reported [11, 12]. Recently, an isoform of GRP78 in the cytosol generated by alternative splicing of the same gene is also reported [13]. GRP78 is contributing in many functions in different parts

of the cell including the plasma membrane. [5, 14-23]. Berger et al reported the cell surface localization of GRP78 for the first time in 1997 [24].

Cancer and ER stress

The metabolic environment of tumors is often acidic, hypoxia, with lack of nutrient and low amounts of both amino acids and glucose [5]. These phenomena occur due to both poor vascularization and rapid growth of tumor cells, and the intrinsic property of cancer cells with high glucose metabolism and higher glycosylation rates. The microenvironment of tumor cells resembles physiologically to ER under stress condition where it causes the UPR is to be activated to help the cells survival [25, 26]. In the case of cancer, the proapoptotic pathways inactivated due to mutations resulting from tumorigenesis, thereby suppressing will be eliminated in the cancerous cells. This, coupled with the activation of the prosurvival UPR pathways, offers an advantage for cancer progression. [27, 28]

Mechanism for GRP78 in promoting cancer progression GRP78 binds and inhibits the activation of caspase-7, an executor caspase activated by both ER stress and genotoxic drugs. GRP78 also binds and suppresses the activation

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of the proapoptotic protein BIK, its downstream target BAX, and preventing release of cytochrome C from the mitochondria. The results obtained from the mouse model showed that Grp78 heterozygosity extremely prolongs the latency and delays the progression of the oncogene-induced mammary tumors in the well-established model without affecting mouse growth, organ development, and antibody production [29].

Based on results reported by Zhou et al. in the MCF-7/BUS-10, a human breast cancer cell line, overexpression of GRP78 suppresses apoptosis induced by BIK and NOXA, alone or in combination. They farther reported that BCL-2 and GRP78 form independent complex with BIK where the increased expression of GRP78 results in decrease BIK binding to BCL-2 [30]. Overexpression of GRP78 also inhibits estrogen starvation-induced BAX activation, mitochondrial permeability transition, and consequent apoptosis in this cell line [31].

Developmental oncoprotein Cripto binds to cell surface and promotes prosurvival signaling GRP78 via MAPK/PI3K and Smad2/3 pathways. Cripto-dependent increase in cellular proliferation is prevented by immunoneutralization of the Cripto-GRP78 complex at the cell surface [32]. Down regulation of MHC-I expression on the cell surface due to overexpression of GRP78 is another mechanism for GRP78 function. As a result of this regulation the capacity of the immune system to control of tumor progression is limited [33]. Interaction of activated 2M (2M*) with GRP78 induces activation of PAK2, ERK1/2, p38 MAPK and P13K thereby promoting cell proliferation [34, 35]. Activation of Akt and NF- B lead to cell survival in prostat cancer. 2M* leads to positive regulation, synthesis and secretion of PSA and a complex between

2M* and PSA serves as a ligand for GRP78. The interaction between GRP78 and this complex further increases the above signaling cascade. Three major mechanisms mediated by GRP78 for cancer progression were suggested by researchers studying on genetic model of breast cancer in the GRP78 heterozygous mice: protection against apoptosis, enhancement of tumor cell proliferation, and promotion of tumor angiogenesis [36-38]. Induction of tumor angiogenesis was approved by another research group. The GRP78 (+/-) mice exhibited a high reduction in the microvessel density (MVD) of the endogenous mammary tumors, where as no effect was observeed on the MVD of normal organs. These observations suggest that the host vasculature is regulated by GRP78 function within the tumor microenvironment [36]. Knockdown of GRP78 significantly suppresses the VEGF-induced activation of ERK1/2, PLC- and VEGF receptor-2 (VEGFR-2).

Consequently the regulatory role of GRP78 in VEGFinduced endothelial cell proliferation through VEGFR-2mediated signalingis also reported [39, 40]. GRP78 has been shown directly to interact with apoptotic pathway intermediates, to block caspase activation, eventually leading to apoptosis inhibition and increased cell survival [31, 41].

Overexpression in cancer

Due to its antiapoptotic properties, GRP78 induction has been reported to be as a prosurvival factor in cells undergoing ER stress. It has been illustrated that GRP78 level is highly elevated in various cancer cell lines, solid associated human cancer biopsies, and has association with malignancy and metastasis [31, 42, 43].

The GRP78 transcription is elevated under various stress conditions suggesting the inVolvement of GRP78 in cell survival enhancement. GRP78 seems to be directly connected to apoptotic pathway intermediates, to block caspase activation which eventually leads to increase cell survival due to apoptosis inhibition [35, 44]. Since tumor progression requires cell proliferation as well as inhibition of tumor cell death, therefore the inherent antiapoptotic properties of GRP78 could play a potential role in cancer progression. This is further supported with the expression level of GRP78, which is markedly higher in primary tumors compared with that of benign tissues. This has been documented in various cancers, including breast, hepatocellular carcinoma [45], lung cancer [46], and prostate cancer [47, 48]. In two different studies Xing and colleagues showed that GRP78 was up- regulated in colon cancer tissue but not in normal tissue. Their results reviled significant increase in GRP78 expression of colorectal carcinoma at the protein level, but there was no difference in the relative mRNA expression levels [49, 50].

Expression of GRP78 in mRNA and protein level was compared between normal primary and esophageal adenocarcinoma tissues. Increased expression of GRP78 in cancer tissue at two levels was related to tumor stage progress [51]. Over expression of GRP78 in glioma was shown by Lee and colleagues. They showed that decreasing of caspase 7 activation and resistance to etoposide and cisplatin-induced apoptosis, is related to the enhancement of GRP78 expression. Using western blot , approximatly 3-fold increase in GRP78 protein level in lung cancer tissue was observed, compared to normal tissue [52].

Induction of chemoresistance

In most of the cases, chemotherapy is the initial strategy for cancer treatment. However, after sometimes cancer grows back without showing response to the initial therapy.GRP78/BiP has been found to be overexpressed, both at the gene and protein levelat this stage. Increased expression of GRP78/BiP in the surviving cells has been found during the treatment of human breast cancer cells MDA-MB-435 with anti-angiogenic factor Combretastatin A4P supporting the correlation of higher GRP78 levels with higher resistance [53]. In addition GRP78 increased expression has been observed in a panel of MCF-7 human breast cancer cell line refractory to several treatments compared to the parental line [54]. Intriguingly, it has been shown that high level of GRP78 expression induced resistance to doxorubicin in breast cancer patients [55, 56]. These observations indicate that GRP78 could be a specific marker to predict doxorubicin resistance in breast cancer.

In addition, Virrey *et al.*, showed that GRP78 induces chemoresistance development in brain endothelial cells and it is believed that the incidence of tumor vascularization and metastatic spread is attributed to this phenomenon [57]. GRP78 inhibition resensitize acute lymphoblastic leukemia cells refractory to vincristine [58]. GRP78 also plays an important role to contribute to castration resistant

prostate cancer (CRPC) development [59]. Immunohistochemical analysis revealed GRP78 overexpression in CRPC and its positive correlation with poor survival recurrence [47].

GRP78 expression has straight relation with the resistance of glioma cells to temozolomide. It has been shown that GRP78 knockdown lowers resistance of cells to temozolomide, where as cells with overexpression of GRP78, confers the higher resistance. Knockdown of GRP78 also sensitizes glioma cells to 5-fluorouracil and CPT-11[60].

Yidan Lin et al. had proposed a role of GRP78 in lung cancer. GRP78 is one of the factors which can regulate phosphorylation of Akt and it is essential to the ER stress-tolerant ERST-associated Cisplatin resistance in lung cancer cells [61].

Tumor treatment with GRP78

GRP78 is highly expressed in various situations such as glucose deprivation, chronic anoxia, and acidic pH, all persisting in poorly vascularized solid tumors. There are reports indicating the importance of GRP78 at different stages of tumorigensis, and variety of reports implicate the potential of GRP78 as a cancer therapeutic target or prognostic factor. In several studies it showed that GRP78 is the target of anticancer agents [62-64].

Either GRP78 expression or its activity is inhibited by pharmacological concentrations of several naturally occurring compounds with putative anticancer activity such as genistein, an active ingredient of soy, (-)– epigallocatechin gallate (EGCG), a green tea component, and salicyclic acid from plants [65-67]. To describe one of these components, EGCG directly interact with GRP78 at its ATP-binding site and regulates its function by competing with ATP binding [65, 68]. On the other hand, some synthetic drugs such as Pyrviniumpamoate, an old anthelmintic medicine, can suppress glucose deprivationinduced GRP78 transcription, and its combination with doxorubicin enhances its antitumor activity [69].

It has been shown that the HKH40A, the 8-methoxy analog of WMC79, promotes it's antitumor activity by reduction of GRP78 protein in cancer cell line[70]. Using GRP78 targeting peptides, when linked to Taxol, induced apoptosis in the targeted cancer cells [71].

Recentluy GRP78 is reported to be present on the surface of tumor cells but not in normal organs which opens up an exciting opportunity of targeting cell surface GRP78 as well as using it as a cancer-targeting marker [72]. Cellular growth suppression and apoptosis induction is occurred by ligation of surface GRP78 with antibody against the C-terminal domain of GRP78 which is exposed exteracellulary. Antibody binding led to reduction of GRP78 expression,up regulation of P53 activity, apoptosis promotion and inhibition of cellular proliferation in melanoma and prostate cell lines [73].

Misra *et al.*, demonstrated that incubation of 1-LN prostate cancer cell with antibody directed towards C-terminal domain of GRP78 led to molecular changes in Ras/MAPK and PI 3-kinase/AKT signaling pathways, reduction of anti appototic Bcl-2 and up regulation of pro-apoptotic BaD, BaX and BaK [74]. Cell death by SAM-6, an anti GRP78 monoclonal antibody, was reported by Brandlein *et al.*,

This IgM antibody binds to glycosylated form of GRP78 leading to apoptosis viainduction of an intrinsic-like form of apoptosis and overaccumulation of large non-physiologic intracellular lipid [75]. Since the targeting of C-terminal domain of GRP78 (CGRP) with antibody is reported as a viable approach for tumor suppression, we evaluated the structure and antigenicity of this domain in previous study [76]. Our results showed that CGRP can play an important role as a target in cancer studies.

The GRP78 promoter-driven transgene is largely quiescent in major adult organs, but highly active in cancer cells and cancer-associated macrophages, which can diffuse to tumor necrotic sites devoid of vascular supply and facilitate cell-based therapy. Thus, controlling the transcription process by using the GRP78 promoter suggests multiple novel approaches for human cancer gene therapy, such as suicide genes, immunosuppressors and tumor suppressors. For example when the GRP78 promoter was used to drive the expression of Herpes simplex virus-thymidine kinase (HSV-tk) suicide gene in a retroviral system, complete eradication of sizable tumor mass, with no recurrence of tumor growth was occurred [77-79].

Conclusion

Together, studies suggest that over expression of GRP78 in cancer cell induces cancer progression and resistance to chemotherapeutic agents. Cell surface expression of GRP78 exclusively on cancer cells is an opportunity for tumor targeting via this protein. For more reliable application of GRP78, some key issues such as the basic mechanism for localization of GRP78 on the cell surface, downstream signaling after ligand binding and how to use this protein as a biological marker in cancer detection must be farther investigated.

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References

1. Shen, X., Zhang, K., Kaufman, R. J., The unfolded protein response--a stress signaling pathway of the endoplasmic reticulum. *J Chem Neuroanat*, 2004, Vol. 28, pp. 79-92.

2. Zhang, K., Kaufman, R. J., The unfolded protein response A stress signaling pathway critical for health and disease. *Neurol*, 2006, Vol. 66, pp. S102-S109.

3. Lee, A.S., The glucose-regulated proteins: stress induction and clinical applications. *Trends Biochem Sci*, 2001, Vol. 26, pp. 504-510.

4. Ma, Y., Hendershot, L. M., The role of the unfolded protein response in tumour development: friend or foe? *Nat Rev Cancer*, 2004, Vol. 4, pp. 966-977.

5. Li, J., Lee, A. S., Stress induction of GRP78/BiP and its role in cancer. *Curr Mol Med*, 2006, Vol. 6, pp. 45-54.

6. Kondo, Y., Kanzawa, T., Sawaya, R., Kondo, S., The role of autophagy in cancer development and response to therapy. *Nat Rev Cancer*, 2005, Vol. 5, pp. 726-734.

7. Bertolotti, A., Zhang, Y., Hendershot, L. M., Harding, H. P., Ron, D., Dynamic interaction of BiP and ER stress transducers in the unfolded-protein response. *Nat Cell Biol*, 2000, Vol. 2, pp. 326-332. 8. TING, J., LEE, A. S., Human gene encoding the 78,000dalton glucose-regulated protein and its pseudogene: structure, conservation, and regulation. *DNA*, 1988, Vol. 7, pp. 275-286.

9. Shen, X., Zhang, K., Kaufman, R. J., The unfolded protein response a stress signaling pathway of the endoplasmic reticulum. *J Chem Neuroanat*, 2004, Vol. 28, pp. 79-92.

10. Munro, S., Pelham, H. R., A C-terminal signal prevents secretion of luminal ER proteins. *Cell*, 1987, Vol. 48, pp. 899-907.

11. Matsumoto, A., Hanawalt, P. C., Histone H3 and heat shock protein GRP78 are selectively cross-linked to DNA by photoactivated gilvocarcin V in human fibroblasts. *Cancer Res*, 2000, Vol. 60, pp. 3921-3926.

12. Sun, F. C., Wei, S., Li, C. W., Chang, Y. S., Chao, C. C., Lai, Y. K., Localization of GRP78 to mitochondria under the unfolded protein response. *Biochem J*, 2006, Vol. 396, pp. 31.

13. Ni, M., Zhou, H., Wey, S., Baumeister, P., Lee, A. S., Regulation of PERK signaling and leukemic cell survival by a novel cytosolic isoform of the UPR regulator GRP78/BiP. *PLoS One*, 2009, Vol. 4, pp. e6868.

14. Arap, M. A., Lahdenranta, J., Mintz, P. J., Hajitou, A., Sarkis, Á. S., Arap, W., Pasqualini, R., Cell surface expression of the stress response chaperone GRP78 enables tumor targeting by circulating ligands. *Cancer Cell*, 2004, Vol. 6, pp. 275-284.

15. Bodman-Smith, M. D., Corrigall, V. M., Kemeny, D.M., Panayi, G.S., BiP, a putative autoantigen in rheumatoid arthritis, stimulates IL-10-producing CD8-positive T cells from normal individuals. *Rheumatol(Oxford)*, 2003, Vol. 42, pp. 637-44.

16. Brownlie, R. J., Myers, L. K., Wooley, P. H., Corrigall, V.M., Bodman-Smith, M. D., Panayi, G. S., Thompson, S. J., Treatment of murine collagen-induced arthritis by the stress protein BiP via interleukin-4–producing regulatory T cells: A novel function for an ancient protein. *Arthritis Rheum*, 2006, Vol. 54, pp. 854-863.

17. Cabrera-Hernandez, A., Thepparit, C., Suksanpaisan, L., Smith, D. R., Dengue virus entry into liver (HepG2) cells is independent of hsp90 and hsp70. *J Med Virol*, 2007, Vol. 79, pp. 386-392.

18. Corrigall, V. M., Bodman-Smith, M. D., Fife, M. S., Canas, B., Myers, L. K., Wooley, P. H., Soh, C., Staines, N. A., Pappin, D. J., Berlo, S. E., The human endoplasmic reticulum molecular chaperone BiP is an autoantigen for rheumatoid arthritis and prevents the induction of experimental arthritis. *J Immunol*, 2001, Vol. 166, pp. 1492-1498.

19. Davidson, D. J., Haskell, C., Majest, S., Kherzai, A., Egan, D.A., Walter, K. A., Schneider, A., Gubbins, E. F., Solomon, L., Chen, Z., Kringle 5 of human plasminogen induces apoptosis of endothelial and tumor cells through surface-expressed glucose-regulated protein 78. *Cancer Res*, 2005, Vol. 65, pp. 4663-4672.

20. Jindadamrongwech, S., Thepparit, C., Smith, D., Identification of GRP 78 (BiP) as a liver cell expressed receptor element for dengue virus serotype 2. *Arch Virol*, 2004, Vol. 149, pp. 915-927.

21. Misra, U. K., Gonzalez-Gronow, M., Gawdi, G., Hart, J. P., Johnson, C. E., Pizzo, S. V., The role of Grp 78 in alpha 2-macroglobulin-induced signal transduction. Evidence from RNA interference that the low density lipoprotein receptor-related protein is associated with, but not necessary for, GRP 78-mediated signal transduction. *J Biol Chem*, 2002, Vol. 277, pp. 42082-7.

22. Panayi, G., Corrigall, V., BiP regulates autoimmune inflammation and tissue damage. *Autoimmun Rev*, 2006, Vol. 5, pp. 140-142.

23. Triantafilou, K., Fradelizi, D., Wilson, K., Triantafilou, M., GRP78, a coreceptor for coxsackievirus A9, interacts with major histocompatibility complex class I molecules which mediate virus internalization. *J Virol*, 2002, Vol. 76, pp. 633-643.

24. Berger, C. L., Dong, Z., Hanlon, D., Bisaccia, E., Edelson, R.L., A lymphocyte cell surface heat shock protein homologous to the endoplasmic reticulum chaperone, immunoglobulin heavy chain binding protein BIP. *Int J Cancer*, 1997, Vol. 71, pp. 1077-1085.

25. Koumenis, C., ER stress, hypoxia tolerance and tumor progression. *Curr Mol Med*, 2006, Vol. 6, pp. 55-69.

26. Bi, M., Naczki, C., Koritzinsky, M., Fels, D., Blais, J., Hu, N., Harding, H., Novoa, I., Varia, M., Raleigh, J., ER stress-regulated translation increases tolerance to extreme hypoxia and promotes tumor growth. *EMBO J*, 2005, Vol. 24, pp. 3470-3481.

27. Fu, Y., Lee, A. S., Glucose regulated proteins in cancer progression, drug resistance and immunotherapy. *Cancer Biol Ther*, 2006, Vol. 5, pp. 741-744.

28. Zhang, J., Jiang, Y., Jia, Z., Li, Q., Gong, W., Wang, L., Wei, D., Yao, J., Fang, S., Xie, K., Association of elevated GRP78 expression with increased lymph node metastasis and poor prognosis in patients with gastric cancer. *Clin Exp Metastas*, 2006, Vol. 23, pp. 401-410.

29. Guy, C., Cardiff, R., Muller, W., Induction of mammary tumors by expression of polyomavirus middle T oncogene: a transgenic mouse model for metastatic disease. *Mol Cell Biol*, 1992, Vol. 12, pp. 954-961.

30. Zhou, H., Zhang, Y., Fu, Y., Chan, L., Lee, A. S., Novel mechanism of anti-apoptotic function of 78-kDa Glucose-Regulated Protein (GRP78) endocrine resistance factor in breast cancer, through release of b-cell lymphoma 2 (bcl-2) from bcl-2-interacting killer (bik). *J Biol Chem*, 2011, Vol. 286, pp. 25687-25696.

31. Fu, Y., Li, J., Lee, A. S., GRP78/BiP inhibits endoplasmic reticulum BIK and protects human breast cancer cells against estrogen starvation–induced apoptosis. *Cancer Res*, 2007, Vol. 67, pp. 3734-3740.

32. Kelber, J.A., Panopoulos, A. D., Shani, G., Booker, E. C., Belmonte, J. C., Vale, W. W., Gray, P. C., Blockade of Cripto binding to cell surface GRP78 inhibits oncogenic Cripto signaling via MAPK/PI3K and Smad2/3 pathways. *Oncogene*, 2009, Vol. 28, pp. 2324-36.

33. Henderson, B., Pockley, A. G., *Cellular Trafficking of Cell Stress Proteins in Health and Disease*. Vol. 6. 2012: Springer.

34. Misra, U. K., Deedwania, R., Pizzo, S. V., Binding of activated 2-macroglobulin to its cell surface receptor GRP78 in 1-LN prostate cancer cells regulates PAK-2-dependent activation of LIMK. *J Biol Chem*, 2005, Vol. 280, pp. 26278-26286.

35. Reddy, R. K., Mao, C., Baumeister, P., Austin, R. C., Kaufman, R.J., Lee, A.S., Endoplasmic reticulum chaperone protein GRP78 protects cells from apoptosis induced by topoisomerase inhibitors: role of ATP binding site in suppression of caspase-7 activation. *J Biol Chem*, 2003, Vol. 278, pp. 20915-24.

36. Dong, D., Ni, M., Li, J., Xiong, S., Ye, W., Virrey, J.J., Mao, C., Ye, R., Wang, M., Pen, L., Dubeau, L., Groshen, S., Hofman, F.M., Lee, A.S., Critical role of the stress chaperone GRP78/BiP in tumor proliferation, survival, and tumor angiogenesis in transgene-induced mammary tumor development. *Cancer Res*, 2008, Vol. 68, pp. 498-505.

37. Misra, U. K., Deedwania, R., Pizzo, S. V., Activation and cross-talk between Akt, NF- B, and unfolded protein response signaling in 1-LN prostate cancer cells consequent to ligation of cell surface-associated GRP78. *J Biol Chem*, 2006, Vol. 281, pp. 13694-13707.

38. Folkman, J., Ingber, D., Inhibition of angiogenesis. *Semin Cancer Biol*, 1992, Vol. 3, pp. 89-96.

39. Katanasaka, Y., Ishii, T., Asai, T., Naitou, H., Maeda, N., Koizumi, F., Miyagawa, S., Ohashi, N., Oku, N., Cancer antineovascular therapy with liposome drug delivery systems

targeted to BiP/GRP78. Int J Cancer, 2010, Vol. 127, pp. 2685-98.

40. Li, Z., Glucose regulated protein 78: a critical link between tumor microenvironment and cancer hallmarks. *Biochim Biophys Acta*, 2012, Vol. 1826, pp. 13-22.

41. Li, J., Ni, M., Lee, B., Barron, E., Hinton, D., Lee, A., The unfolded protein response regulator GRP78/BiP is required for endoplasmic reticulum integrity and stress-induced autophagy in mammalian cells. *Cell Death Differ*, 2008, Vol. 15, pp. 1460-1471.

42. Fu, Y., Lee, A. S., Glucose regulated proteins in cancer progression, drug resistance and immunotherapy. *Cancer Biol Ther*, 2006, Vol. 5, pp. 741-4.

43. Li, J., Lee, A. S., Stress induction of GRP78/BiP and its role in cancer. *Curr Mol Med*, 2006, Vol. 6, pp. 45-54.

44. Fu, Y., Li, J., Lee, A. S., GRP78/BiP inhibits endoplasmic reticulum BIK and protects human breast cancer cells against estrogen starvation-induced apoptosis. *Cancer Res*, 2007, Vol. 67, pp. 3734-40.

45. Shuda, M., Kondoh, N., Imazeki, N., Tanaka, K., Okada, T., Mori, K., Hada, A., Arai, M., Wakatsuki, T., Matsubara, O., Yamamoto, N., Yamamoto, M., Activation of the ATF6, XBP1 and grp78 genes in human hepatocellular carcinoma: a possible inVolvement of the ER stress pathway in hepatocarcinogenesis. *J Hepatol*, 2003, Vol. 38, pp. 605-14.

46. Uramoto, H., Sugio, K., Oyama, T., Nakata, S., Ono, K., Yoshimastu, T., Morita, M., Yasumoto, K., Expression of endoplasmic reticulum molecular chaperone Grp78 in human lung cancer and its clinical significance. *Lung Cancer*, 2005, Vol. 49, pp. 55-62.

47. Daneshmand, S., Quek, M. L., Lin, E., Lee, C., Cote, R. J., Hawes, D., Cai, J., Groshen, S., Lieskovsky, G., Skinner, D.G., Lee, A.S., Pinski, J., Glucose-regulated protein GRP78 is upregulated in prostate cancer and correlates with recurrence and survival. *Hum Pathol*, 2007, Vol. 38, pp. 1547-52.

48. Pootrakul, L., Datar, R. H., Shi, S. R., Cai, J., Hawes, D., Groshen, S. G., Lee, A. S., Cote, R.J., Expression of stress response protein Grp78 is associated with the development of castration-resistant prostate cancer. *Clin Cancer Res*, 2006, Vol. 12, pp. 5987-93.

49. Xing, X., Lai, M., Wang, Y., Xu, E., Huang, Q., Overexpression of glucose-regulated protein 78 in colon cancer. *Clin Chim Acta*, 2006, Vol. 364, pp. 308-315.

50. Xing, X., Li, Y., Liu, H., Wang, L., Sun, L., Glucose regulated protein 78 (GRP78) is overexpressed in colorectal carcinoma and regulates colorectal carcinoma cell growth and apoptosis. *Acta Histochem*, 2011, Vol. 113, pp. 777-782.

51. Langer, R., Feith, M., Siewert, J. R., Wester, H. J., Hoefler, H., Expression and clinical significance of glucose regulated proteins GRP78 (BiP) and GRP94 (GP96) in human adenocarcinomas of the esophagus. *BMC cancer*, 2008, Vol. 8, pp. 70.

52. Wang, Q., He, Z., Zhang, J., Wang, Y., Wang, T., Tong, S., Wang, L., Wang, S., Chen, Y., Overexpression of endoplasmic reticulum molecular chaperone GRP94 and GRP78 in human lung cancer tissues and its significance. *Cancer Detect Prev*, 2005, Vol. 29, pp. 544-551.

53. Dong, D., Ko, B., Baumeister, P., Swenson, S., Costa, F., Markland, F., Stiles, C., Patterson, J. B., Bates, S.E., Lee, A. S., Vascular targeting and antiangiogenesis agents induce drug resistance effector GRP78 within the tumor microenvironment. *Cancer Res*, 2005, Vol. 65, pp. 5785-91.

54. Wosikowski, K., Schuurhuis, D., Kops, G. J., Saceda, M., Bates, S. E., Altered gene expression in drug-resistant human breast cancer cells. *Clin Cancer Res*, 1997, Vol. 3, pp. 2405-14.

55. Lee, E., Nichols, P., Spicer, D., Groshen, S., Yu, M. C., Lee, A. S., GRP78 as a novel predictor of responsiveness to

chemotherapy in breast cancer. Cancer Res, 2006, Vol. 66, pp. 7849-53.

56. Scriven, P., Coulson, S., Haines, R., Balasubramanian, S., Cross, S., Wyld, L., Activation and clinical significance of the unfolded protein response in breast cancer. *Br J Cancer*, 2009, Vol. 101, pp. 1692-8.

57. Virrey, J. J., Dong, D., Stiles, C., Patterson, J. B., Pen, L., Ni, M., Schonthal, A. H., Chen, T. C., Hofman, F. M., Lee, A. S., Stress chaperone GRP78/BiP confers chemoresistance to tumor-associated endothelial cells. *Mol Cancer Res*, 2008, Vol. 6, pp. 1268-75.

58. Uckun, F. M., Qazi, S., Ozer, Z., Garner, A. L., Pitt, J., Ma, H., Janda, K. D., Inducing apoptosis in chemotherapy-resistant B-lineage acute lymphoblastic leukaemia cells by targeting HSPA5, a master regulator of the anti-apoptotic unfolded protein response signalling network. *Br J Haematol*, 2011, Vol. 153, pp. 741-52.

59. Tan, S. S., Ahmad, I., Bennett, H. L., Singh, L., Nixon, C., Seywright, M., Barnetson, R. J., Edwards, J., Leung, H. Y., GRP78 up-regulation is associated with androgen receptor status, Hsp70-Hsp90 client proteins and castrate-resistant prostate cancer. *J Pathol*, 2011, Vol. 223, pp. 81-7.

60. Pyrko, P., Schönthal, A. H., Hofman, F. M., Chen, T. C., Lee, A.S., The unfolded protein response regulator GRP78/BiP as a novel target for increasing chemosensitivity in malignant gliomas. *Cancer Res*, 2007, Vol. 67, pp. 9809-9816.

61. Lin, Y., Wang, Z., Liu, L., Chen, L., Akt is the downstream target of GRP78 in mediating cisplatin resistance in ER stress-tolerant human lung cancer cells. *Lung Cancer*, 2011, Vol. 71, pp. 291-297.

62. Luo, B., Lee, A. S., The critical roles of endoplasmic reticulum chaperones and unfolded protein response in tumorigenesis and anticancer therapies. *Oncogene*, 2013, Vol. 32, pp. 805-818.

63. Schwarze, S., Rangnekar, V., Targeting plasma membrane GRP78 for cancer growth inhibition. *Cancer Biol Ther*, 2010, Vol. 9, pp. 153-155.

64. Lee, A. S., Glucose-regulated proteins in cancer: molecular mechanisms and therapeutic potential. *Nat Rev Cancer*, 2014, Vol. 14, pp. 263-276.

65. Ermakova, S. P., Kang, B. S., Choi, B. Y., Choi, H. S., Schuster, T. F., Ma, W. Y., Bode, A. M., Dong, Z., (–)– Epigallocatechin Gallate Overcomes Resistance to Etoposide-Induced Cell Death by Targeting the Molecular Chaperone Glucose-Regulated Protein 78. *Cancer Res*, 2006, Vol. 66, pp. 9260-9269.

66. Zhou, Y., Lee, A. S., Mechanism for the suppression of the mammalian stress response by genistein, an anticancer phytoestrogen from soy. *J Natl Cancer Inst*, 1998, Vol. 90, pp. 381-388.

67. Deng, W. G., Ruan, K. H., Du, M., Saunders, M. A., Wu, K. K., Aspirin and salicylate bind to immunoglobulin heavy chain binding protein (BiP) and inhibit its ATPase activity in human fibroblasts. *FASEB J*, 2001, Vol. 15, pp. 2463-2470.

68. Li, M., Wang, J., Jing, J., Hua, H., Luo, T., Xu, L., Wang, R., Liu, D., Jiang, Y., Synergistic promotion of breast cancer cells death by targeting molecular chaperone GRP78 and heat shock protein 70. *J Cell Mol Med*, 2009, Vol. 13, pp. 4540-4550.

69. Yu, D. H., Macdonald, J., Liu, G., Lee, A. S., Ly, M., Davis, T., Ke, N., Zhou, D., Wong-Staal, F., Li, Q. X., Pyrvinium targets the unfolded protein response to hypoglycemia and its anti-tumor activity is enhanced by combination therapy. *PLoS One*, 2008, Vol. 3, pp. e3951.

70. Kosakowska-Cholody, T., Lin, J., Srideshikan, S., Scheffer, L., Tarasova, N., Acharya, J., HKH40A downregulates GRP78/BiP expression in cancer cells. *Cell Death Dis*, 2014, Vol. 5, pp. e1240.

71. Kim, Y., Lillo, A. M., Steiniger, S. C., Liu, Y., Ballatore, C., Anichini, A., Mortarini, R., Kaufmann, G.F., Zhou, B., Felding-Habermann, B., Targeting heat shock proteins on cancer cells: selection, characterization, and cell-penetrating properties of a peptidic GRP78 ligand. *Biochem*, 2006, Vol. 45, pp. 9434-9444. 72. Zhang, Y., Liu, R., Ni, M., Gill, P., Lee, A. S., Cell surface relocalization of the endoplasmic reticulum chaperone and unfolded protein response regulator GRP78/BiP. *J Biol Chem*, 2010, Vol. 285, pp. 15065-15075.

73. Misra, U. K., Mowery, Y., Kaczowka, S., Pizzo, S.V., Ligation of cancer cell surface GRP78 with antibodies directed against its COOH-terminal domain up-regulates p53 activity and promotes apoptosis. *Mol Cancer Ther*, 2009, Vol. 8, pp. 1350-62. 74. Misra, U. K., Pizzo, S. V., Ligation of cell surface GRP78 with antibody directed against the COOH-terminal domain of GRP78 suppresses Ras/MAPK and PI 3-kinase/AKT signaling while promoting caspase activation in human prostate cancer cells. *Cancer Biol Ther*, 2010, Vol. 9, pp. 142-52

75. Brändlein, S., Rauschert, N., Rasche, L., Dreykluft, A., Hensel, F., Conzelmann, E., Müller-Hermelink, H. K., Vollmers, H.P., The human IgM antibody SAM-6 induces tumor-specific apoptosis with oxidized low-density lipoprotein. *Mol cancer Ther*, 2007, Vol. 6, pp. 326-333.

76. Aghamollaei, H., Gargari, S. L. M., Ghanei, M., Rasaee, M. J., Amani, J., Bakherad, H., Farnoosh, G., Structure prediction, expression and antigenicity of c-terminal of GRP78. *Biotechnol Appl Biochem*, 2015, In press.

77. Gazit, G., Hung, G., Chen, X., Anderson, W.F., Lee, A.S., Use of the glucose starvation-inducible glucose-regulated protein 78 promoter in suicide gene therapy of murine fibrosarcoma. *Cancer Res*, 1999, Vol. 59, pp. 3100-3106.

78. Chen, X., Zhang, D., Dennert, G., Hung, G., Lee, A. S., Eradication of murine mammary adenocarcinoma through HSVtk expression directed by the glucose-starvation inducible grp78 promoter. *Breast cancer Res Treat*, 2000, Vol. 59, pp. 81-90.

79. Azatian, A., Yu, H., Dai, W., Schneiders, F. I., Botelho, N. K., Lord, R. V., Effectiveness of HSV-tk suicide gene therapy driven by the Grp78 stress-inducible promoter in esophagogastric junction and gastric adenocarcinomas. *J Gastrointest Surg*, 2009, Vol. 13, pp. 1044-1051.