

Hepatitis C among Hemodialysis Patients: A Review on Epidemiologic, Diagnostic, and Therapeutic Features

Seyed-Moayed Alavian ¹, Seyed Mohammad-Mehdi Hosseini-Moghaddam ^{2*}, Mohammad Rahnavardi ²

¹ Baqiyatallah Research Center for Gastroenterology and Liver Diseases, Baqiyatallah University of Medical Sciences & Tehran Hepatitis Center, Tehran, Iran

² Urology and Nephrology Research Center (UNRC), Shaheed Beheshti University of Medical Sciences, Tehran, Iran

Hepatitis C virus (HCV) is a major public health problem and is the most common liver disease among hemodialysis (HD) patients. The seroprevalence of HCV infection among HD ranged from 1.9% to 80% in reports published since 1999. The main risk factor for HCV acquisition in HD patients seems the length of time on HD. Phylogenetic analysis of HCV viral isolates has suggested nosocomial patient-to-patient transmission of HCV infection among HD patients. Lack of strict adherence to universal precautions by staff and sharing of articles such as multidose drugs might be the main mode of nosocomial HCV spread among HD patients. Currently, there are several dilemmas on the management of these patients: should HCV-RNA testing be included in the routine screening of HD population for HCV infection?; does periodic serum alanine aminotransferase testing have a role in screening HD patients for HCV infection?; can dialysis really 'save' the liver of HCV-infected HD patients?; should HCV-infected subjects be isolated and dialyzed by segregated machines?; is there any difference in treating HD and non-HD HCV-infected subjects? This article gathers the present evidence to address these issues and to demonstrate the current worldwide magnitude of HCV in HD population.

Keywords: Hepatitis C Virus, Hemodialysis, Genotype

Introduction

Hepatitis C virus (HCV), infecting about 170 million persons worldwide, is a major public health problem ⁽¹⁾. An estimated 5-20% of HCV-infected patients have or will develop cirrhosis, 1-4% of whom will annually develop hepatocellular carcinoma. Well-known risk factors for HCV transmission include injection drug use, blood product transfusion, organ transplantation, chronic hemodialysis (HD), occupational exposure among health care workers, unprotected sexual contact, and vertical transmission ^(2, 3).

The relation between HCV infection and kidney disorders is well recognized. Hepatitis C infection has been associated with essential mixed cryoglobulinemia that may lead to

membranoproliferative glomerulonephritis ⁽⁴⁾. On the other hand, patients with renal disease have been at increased risk of acquiring HCV because of prolonged vascular access as well as the potential for exposure to infected patients and contaminated

* Correspondence:

Seyed Mohammad-Mehdi Hosseini-Moghaddam MD, MPH, Urology and Nephrology Research Center (UNRC), No.44, 9th Boushan, Pasdaran Avenue, Tehran, Iran.

Tel: +98 21 22567222

Fax: +98 21 22567282

E-mail: h_sasan@hotmail.com

Received: 20 Jun 2007

Revised: 6 Oct 2007

Accepted: 11 Oct 2007

Hep Mon 2007; 7 (3): 153-162

equipment. Liver disease is a significant cause of morbidity and mortality in patients with end-stage renal disease (ESRD) treated by dialysis or transplantation and hepatitis C is the most common liver disease in renal dialysis patients⁽⁵⁾. This article is an update on the current worldwide magnitude of HCV in ESRD patients under HD, the diagnostic features, natural history, preventive measures, and current therapeutic advances in this particular population.

Historical Aspects and Virology

In 1974, Prince *et al.*⁽⁶⁾ firstly reported non-A, non-B viral hepatitis. Fifteen years later, Choo *et al.*⁽⁷⁾ discovered and described HCV. HCV is a small RNA virus that is included in the Flaviviridae family and has been recently classified as the sole member of the Hepacivirus genus⁽⁸⁾. HCV isolates are classified into 6 major genotypes and more than 50 subtypes⁽⁹⁾.

HCV Global Epidemiologic Features in HD Population

The prevalence of HCV infection varies greatly among various populations of patients on HD from different geographic regions. In a review of so far published data in 1999, Wreghitt⁽¹⁰⁾ described HCV prevalence among HD population ranging from 4% in the UK to 71% in Kuwait. Some investigators suggested a decline in HCV prevalence among HD patients in recent years, mostly attributable to strict adherence to universal precautions and with⁽¹¹⁻¹⁷⁾ or without^(18, 19) observing isolation measures.

The reported anti-HCV positivity since 1999 ranged from 1.9% in the Slovenian 2001 annual report⁽²⁰⁾ to 80% in Senegal⁽²¹⁾. The HCV seroprevalence in HD population was 59% in Bosnia and Herzegovina⁽²²⁾, 6.8% in Belgium⁽¹¹⁾, 16.3% in France⁽²³⁾, 6.1% in Germany⁽²⁴⁾, 10-29% in Greece⁽²⁵⁻²⁷⁾, 22.5-32.1% in Italy^(28, 29), 75% in Moldavia⁽³⁰⁾, 3.4% in the Netherlands⁽³¹⁾, 11% in Sweden⁽³²⁾, 7-23.3% in the USA⁽³³⁻³⁷⁾, 20.5% in Libya⁽³⁸⁾, 80% in Senegal⁽²¹⁾, 23.7% in Sudan⁽³⁹⁾, 19-41.7% in Tunisia^(40, 41), 8.4-43.2% in Brazil^(42, 43), 6.7% in Mexico⁽⁴⁴⁾, 59.3% in Peru⁽⁴⁵⁾, and 3.5% in Puerto Rico⁽⁴⁶⁾.

Table 1 describes the results of reports from Asian countries since 2000 on HCV seroprevalence among HD patients. The studies that prospectively

Table 1. Reports on HCV seroprevalence among HD patients in Asian countries (2000-2007).

Study	Country	Year	HD centers enrolled	HCV seropositivity/Patients (%)
Hosseini-Moghaddam <i>et al.</i> ⁽⁶⁴⁾	Iran	2006	45	155/1914 (8.1)
Amiri <i>et al.</i> ⁽⁵⁵⁾	Iran	2005	7	80/298 (24.8)
Alavian <i>et al.</i> ⁽⁵⁶⁾	Iran	2003	26	111/838 (13.2)
Ansar <i>et al.</i> ⁽⁵⁷⁾	Iran	2002	1	52/93 (55.9)
Furusyo <i>et al.</i> ⁽⁵¹⁾	Japan	2001	1	100/269 (37.2)
Iwasaki <i>et al.</i> ⁽⁹³⁾	Japan	2000	1	34/142 (23.9)
Bdour <i>et al.</i> ⁽⁵⁸⁾	Jordan	2002	6	98/283 (34.6)
Hussein <i>et al.</i> ⁽⁵⁹⁾	Saudi Arabia	2007	1	34/180 (18.9)
Al-Shohaib <i>et al.</i> ⁽⁶⁰⁾	Saudi Arabia	2003	3	73/139 (52.5)
Othman <i>et al.</i> ⁽⁶²⁾	Syria	2001	2	68/139 (48.9)
Ocak <i>et al.</i> ⁽⁶⁵⁾	Turkey	2006	3	34/267 (12.7)
Harmankaya <i>et al.</i> ⁽¹⁰²⁾	Turkey	2002	1	8/168 (4.7)

followed HD patients for their HCV status presented an annual incidence rate of de novo HCV infection of 0.4% in France⁽⁴⁷⁾, 0.5% in Tunisia⁽⁴⁷⁾, 0.5% in the Netherlands⁽³¹⁾, 0.83% in Italy⁽²⁸⁾, 1.38%⁽⁴⁸⁾ and 2.1%⁽⁴⁹⁾ in the USA, 0.33%⁽⁵⁰⁾, 2.59%⁽⁵¹⁾, and 3.1% in Japan⁽⁵²⁾, 3.7%⁽⁵³⁾ and 5.5% in Brazil⁽⁵⁴⁾, and 6.2% in Greece⁽²⁷⁾.

Almost all recent surveys on the subject have congruently suggested the length of time on HD as a risk factor for HCV seropositivity^(22-24, 27, 30, 31, 35, 37, 39, 42, 43, 52, 55-61). Historically, the number of blood transfusions received was consistently reported in the literature to be associated with increased prevalence of HCV positive dialysis patients⁽¹⁰⁾. However, several recent reports could not recognize blood transfusion as an independent risk factor in HCV spread among HD subjects^(23, 27, 31, 35, 39, 55, 60-63). Indeed, erythropoietin prescription from the late 1980s onward reduced the HD patients' need to blood transfusion. Furthermore, introduction of sensitive tests for the screening of blood donors has markedly reduced the risk of HCV transmission through blood product transfusion. These two main reasons may explain recent findings on the lack of association between blood transfusion and HCV infection. History of organ transplantation^(23, 27, 31, 55), older age^(17, 39, 48), younger age⁽³⁷⁾, dialysis in multiple centers^(29, 39, 43, 64), the HD unit⁽⁵⁴⁾, hepatitis B infection^(23, 52), human immunodeficiency virus infection^(48, 52), and diabetes mellitus^(17, 65) were other factors that were suggested by some studies to be associated with HCV positivity.

Diagnostic Features

In non-HD population, HCV antibody testing by an enzyme-linked immunosorbent assay (ELISA) is generally used as a screening tool and recombinant immunoblot assay (RIBA) is considered as a confirmatory test because of its high specificity⁽⁶⁶⁻⁶⁸⁾. Viral-based testing is widely accepted as the gold standard in HCV detection. HCV-RNA testing is essential for confirmation of active HCV infection and for monitoring of antiviral therapy. Both qualitative and quantitative tests for HCV-RNA have been developed recently; although, the sensitivities of quantitative tests are lower than qualitative PCR assays⁽⁶⁹⁻⁷¹⁾.

Routine serological testing for HCV infection among HD patients is currently recommended^(72, 73). Current recommendations of Centers for Disease Control and Prevention (CDC) for HCV screening in HD patients include anti-HCV and serum alanine aminotransferase (ALT) testing on admission, monthly ALT, and semiannual anti-HCV^(72, 73). However, the cost-effectiveness of such an approach is under debate. Saab *et al.*⁽⁶⁹⁾ demonstrated that serological-based screening (anti-HCV testing) is less costly and more effective than biochemical-based screening (ALT plus anti-HCV testing) in the diagnosis of *de novo* HCV infection in HD subjects. In comparison to healthy individuals, serum aminotransferase levels are depressed in patients with chronic renal failure (CRF) not requiring dialysis and aspartate aminotransferase (AST) and ALT activity are even lower in dialysis than predialysis patients with CRF⁽⁷⁴⁾. A newly elevated aminotransferase level was found to be neither sensitive nor positively predictive for chronic HCV infection⁽³⁶⁾. However, Fabrizi *et al.*⁽⁴⁹⁾ found that ALT level rose into the abnormal range in newly HCV-infected HD patients and thus suggested the need to monitor chronic HD patients by serial ALT testing.

A dilemma exists on the value of serology since some investigators reported a high rate of false-negative serologic testing^(25, 34). However, the current literature reflects conflicting results in the topic since the frequency of HCV-RNA positive, anti-HCV negative HD patients ranged from 0-12% of all studied HD subjects from several recent reports^(25, 26). A study in India presented a high proportion of HCV-RNA positive, anti-HCV negative subjects (30/124; 24.2%) among the studied CRF population treated with HD or renal transplantation⁽⁷⁵⁾.

The immunocompromised state of HD patients is usually regarded as an explanation for their

deficient antibody response to HCV virus^(5, 73). On the other hand, the reported figures for false-negative serology in some studies might be an overestimate because follow-up samples to detect possible antibody seroconversions were not obtained on these patients⁽⁷³⁾. A relatively large study in 562 HD patients showed that the median numbers of days that the HCV-RNA assay detected HCV infection earlier than anti-HCV testing was 246 and 154 days for the second and third generation ELISA, respectively⁽²⁷⁾. Considering the results of large studies showing only 5/1323 (0.38%)⁽²³⁾, 24/2777 (0.8%)⁽²⁴⁾, and 2/2286 (0.1%)⁽³¹⁾ and some^(26, 33, 42, 51, 76) without any false-negative serology, routine testing for HCV-RNA to diagnose HCV infection in the HD patients seems unjustified. Congruently, the latest CDC guideline does not recommend the use of reverse transcriptase polymerase chain reaction (RT-PCR) for HCV-RNA detection as the primary test for routine screening. Nonetheless, RT-PCR should still be considered as a confirmatory test when the patient tests positive for anti-HCV or if ALT levels are persistently abnormal in those who are anti-HCV negative in the absence of another etiology⁽⁷³⁾. It is also noteworthy that a single negative anti-HCV test cannot rule out HCV infection in HD population because of the potential latency between infection and seroconversion as well as possible lower sensitivity of ELISA in HD patients as discussed above.

Recent advance in diagnosing early HCV infection is made by detecting the HCV core antigen (HCVcAg) that is present during the early stage of infection when anti-HCV seroconversion has not established. The strong point of this technique is the relative ease of performing ELISA for HCVcAg than assays for HCV-RNA based on gene technology. Additionally, HCVcAg testing permits the detection of an HCV infection about 1.5 months earlier than the HCV antibody screening tests and an average of only 2 days later than quantitative HCV-RNA detection in individual specimens⁽⁷⁷⁾. In relation to the results obtained with the amplicor HCV monitor test, Fabrizi *et al.*⁽⁷⁴⁾ calculated the efficacy of HCVcAg ELISA to be 95.9%, with a sensitivity of 92.7%, a specificity of 97.4%, and positive and negative predictive values of 94.7% and 96.5%, respectively. In one study⁽⁴⁰⁾, there were no HCV-RNA positive patients who tested negative in both HCVcAg and anti-HCV antibody. HCVcAg ELISA would be useful in screening asymptomatic HCV carriers or *de novo* infection in HD patients. Combination of anti-HCV antibody and HCVcAg ELISA assays

would add the sensitivity of the screening program. Since the concentrations of HCV core Ag and HCV-RNA levels are significantly correlated^(40, 74, 78), a further presented advantage of HCVcAg detection could be its role as a reliable marker of HCV replication in anti-HCV positive patients. Thus, it could help in the diagnosis of active HCV infection in anti-HCV positive therapy-naïve individuals, especially in poor-resource settings^(40, 74).

Natural History of HCV in HD Patients

Evaluating the natural history of HCV infection among HD patients faces great controversy because the onset is rarely recognized; the course of HCV is usually indolent and extends over decades rather than years; and HD patients may actually die from various co-morbid conditions before the long-term consequences of HCV infection establishes. Severity of histological changes and HCV-RNA levels were not associated in several series⁽⁷⁹⁻⁸³⁾ and ALT level alone could not predict the extent of the liver damage of HD patients with HCV viremia. HCV-infected HD patients may develop liver damage despite normal ALT levels^(79, 84). Therefore, liver biopsy is the only accurate means of assessing the severity of the hepatitis C infection. The frequency of bridging hepatic fibrosis or cirrhosis ranged from 5% to 32% in various series of HCV-infected HD patients⁽⁸⁵⁾.

Several studies reported the disease activity in HCV-infected HD patients to be mild to moderate and usually milder than non-HD subjects^(80, 81, 86-88). There are several explanations for this phenomenon including the altered immunologic state of the ESRD patients under HD, the relatively low HCV viral load in the HD population with HCV infection⁽⁸⁹⁾ probably secondary to the clearance of HCV RNA by the dialysate and/or the entrapment of HCV-RNA particles onto the membrane surface of dialyzers, marked and prolonged hepatocyte growth factor (HGF) release in HD compared to non-HD HCV-infected subjects⁽⁸⁶⁾ regarding the suggested acceleration in the liver regeneration by exogenous HGF administration in animal studies, and marked endogenous interferon (IFN) alpha increment after HD using both cellulosic and synthetic membranes⁽⁹⁰⁾ which can contribute to reduction in HCV viremia. The issue deserves further studies in larger series of patients before the actual role of dialysis in "saving" liver from hepatitis can be confirmed.

Several well-designed prospective studies aimed to address the natural history of HCV infection in HD

population including the patients' survival. In an important multi-centric prospective study from Japan, Nakayama *et al.*⁽⁹¹⁾ followed up 1470 HD patients (276 positive anti-HCV patients) from 16 dialysis centers for an average of 6 years. Mortality was significantly higher in the anti-HCV-positive than -negative subjects (33% versus 23%). Hepatocellular carcinoma (5.5% versus 0.0%) and liver cirrhosis (8.8% versus 0.4%) were significantly more frequent causes of death in anti-HCV-positive than -negative patients. They presented anti-HCV positivity as a risk factor for death with an adjusted relative risk of 1.57 (95% CI: 1.23-2.00). In another study based on a US national database of 13664 HD patients, Kalantar-Zadeh *et al.*⁽³⁷⁾ reported a significant 1.25 (95% CI: 1.12 to 1.39) mortality hazard ratio for HCV infection. Fabrizi *et al.*⁽⁹²⁾ performed a meta-analysis and introduced the summary estimate for relative risk of HCV infection on mortality in HD patients to be 1.57 (95% CI: 1.33-1.86), a rate close to that of Nakayama *et al.*⁽⁹¹⁾ probably because they enrolled this large study in their meta-analysis. Since the frequency of hepatocellular carcinoma and liver cirrhosis as causes of death was significantly higher among anti-HCV-positive than -negative HD patients in all enrolled surveys, the investigators of this meta-analysis suggested that the increased mortality in anti-HCV-positive patients was at least partially related to chronic liver disease with its attendant complications.

HCV Nosocomial Transmission and Preventive Strategies

Nosocomial patient-to-patient transmission of HCV infection among HD patients is suggested by several investigators who performed phylogenetic analysis of HCV viral isolates^(31, 32, 47, 48, 76, 93-98). Lack of strict adherence to universal precautions by staff and sharing of articles such as multidose drugs might be the main mode of nosocomial HCV spread among HD patients^(48, 93, 97-101). Although some studies found that nosocomial spread of HCV declined when HCV infected patients were treated in dedicated HD units^(12-17, 102, 103), other investigators could control nosocomial spread of HCV among HD patients by strict application of hygienic precautions without isolation of HCV-infected subjects or machine segregation^(18, 100, 104). Indeed, the presented efficacy of isolation might be simply due to the prevented sharing of articles between patients and might reflect a better implementation of other hygienic precautions.

Thereupon, in the absence of more convincing evidence, isolation of HCV-infected dialysis patients and use of dedicated machines are currently unjustified (73, 105). Strict adherence to universal precautions seems to be enough to control disease spread in HD units. CDC recommends especial precautions to be observed in dialysis units including the wearing and changing of gloves and water-proof gowns between patients, systematic decontamination of the equipment, circuit, and surfaces after each patient treatment, no sharing of instruments (e.g., tourniquets) or medications (e.g., multiuse vials of heparin) among patients, and the assignment of patients to specific HD units (73).

Therapeutic Aspects

Treatment of HCV-infected non-uremic patients aims at slowing down the disease progression as well as preventing the hepatic and extrahepatic complications (3, 106). Several outcome measures are used to evaluate response to treatment: sustained virologic response (SVR), the index of HCV eradication, is achieved when HCV-RNA is not detected in serum at the end of treatment and 6 months later by a sensitive test; end of treatment response (ETR) is the continued undetectable virus at completion of treatment; and early virologic response (EVR) is a 2-log drop or loss of HCV-RNA, 12 weeks into therapy.

Since 1991 when IFN- α was approved for use in the treatment of HCV infection, several therapeutic advances have been achieved. Thereafter, experiencing combination therapy with ribavirin and IFN was a major breakthrough that improved the SVR rate to as high as 43% (107). The current optimal HCV treatment also includes IFN-based regimens together with antiviral therapy (3, 108). However, introduction of a new formulation of IFN, namely pegylated IFN (peginterferon) (PEG-IFN) was another major advance in HCV treatment. Pegylation is the binding of the inert polyethylene glycol moiety to IFN molecule that decreases renal clearance despite retained biological activity and makes once-weekly administration of the drug possible (109). Two different PEG-IFN formulations are available now, namely PEG-IFN α -2a (Pegasys) and PEG-IFN α -2b (Peg-Intron). PEG-IFN, when used in combination with ribavirin, conferred a relatively high SVR rate of 54% using PEG-IFN α -2b (110) and 56% using PEG-IFN α -2a (111). Since SVR for genotype 2 or 3 infection was similar among patients treated for 24 versus 48 weeks, 24 weeks of treatment is usually

considered sufficient to treat HCV infection with genotype 2 or 3 (111). In contrast, the 48-week regimen should be used for those infected with HCV genotype 1 because HCV infection with this genotype was found to be the strongest negative predictor of response to PEG-IFN α and ribavirin (110, 112). The goals of therapy in HCV-infected HD patients are not different from those of non-HD ones, as were described above. Importantly, the pre-transplant period in a HCV-infected HD patient should be considered as the 'golden' time for HCV treatment in this population. The main reason is that at present there is no safe and efficient therapy for HCV infection after kidney transplantation (113-115).

Similar to immunocompetent patients, IFN-based treatment for chronic hepatitis C is the mainstay therapy in HD population. Two meta-analyses showed that IFN monotherapy was even more effective in HD than non-uremic patients (116, 117), probably because of decreased IFN clearance rate in uremic patients (118). Nonetheless, more adverse events in this population than non-uremic patients as well as marked estimated mean dropout rates of 17% (117) and 29.6% (116) were reported. Neurological (21%), flu-like (17%), and gastrointestinal (18%) symptoms were the most frequent side effects requiring interruption of treatment in a meta-analysis (117). The gathered data in the current review shows that standard IFN α monotherapy in chronic HD patients have resulted in SVR rates ranged from 18.9% to 58.6% in different studies. The two meta-analyses estimated the overall mean of SVR rate to be 39% (117) and 33% (116). Despite the initial promising SVR rates, not all HCV-positive HD patients should be treated with IFN because the risk-benefit ratio of administration of IFN is not well-known. It is reasonable to consider IFN therapy for all HCV-infected HD patients waiting for a kidney transplant because no post-transplant HCV relapse has occurred after successful pre-transplant IFN α therapy in several series (119-121). IFN therapy for other HD patients should be justified according to their life expectancy, histologic severity of liver disease, and comorbidities. IFN α should be administered 3 million units, subcutaneously, trice weekly preferably for 48 weeks regardless of HCV genotype, because the 48-week regimen was associated with less relapses than the 24-week regimen (121, 122).

ESRD patients have reduced clearance of ribavirin, a drug with mainly renal metabolism. High ribavirin serum level poses a high risk of severe hemolytic anemia in ESRD patients, who are often

already anemic. Hence, ribavirin is contraindicated in HD subjects. However, Tan *et al.* (123) used combination IFN α -2b and low-dose ribavirin (200 mg/day) in five patients. Ribavirin was stopped permanently in two patients because of severe anemia. Safety and efficacy of this combination therapy should be studied in larger trials before any recommendation can be made.

In immunocompetent patients, as discussed above, the current optimal treatment for HCV infection includes the combination of PEG-IFN and ribavirin. A pharmacokinetic study suggested the absorption, distribution, and body clearance of PEG-IFN α -2a were not significantly different in subjects with normal renal function versus renal failure, non-dialysis-dependent patients (124). Although this pharmacokinetic study suggests that the use of PEG-IFN in HD population might be possible with comparable safety to non-uremic patients, one relatively large clinical study, treating 78 HCV-infected HD subjects with PEG-IFN α -2a (135 μ g weekly), reported a high dropout frequency of 32% while 83% of patients experienced adverse events. The SVR was 14.1% in this study (125). A randomized trial on either 1.0 or 0.5 μ g/kg of PEG-IFN α -2b initially enrolled 16 patients but was terminated because of adverse events. After modification to the study design, the trial continued, yet 7/16 (44%) required discontinuation of therapy. Only 2 subjects in the 1.0 μ g/kg (22%) and none in the 0.5 μ g/kg groups showed SVR. Whereas these investigators could not find better results of PEG-IFN therapy compared to other reports from administration of IFN regimen, they observed poor tolerance and substantial side effects of the used pegylated formulation (126). Nevertheless, Kokoglu *et al.* (127) administered PEG-IFN α -2a at a dose of 135 μ g weekly for 48 weeks in 12 HCV-infected HD patients and reported no dropouts despite frequent side effects (anemia in 75%, fatigue in 58%, thrombocytopenia in 33%, and leucopenia in 33%). SVR rate was surprisingly as high as 75% (9/12) in this study. In another trial, Sporea *et al.* (128) observed SVR in 3/10 (30%) patients with 4/10 discontinued therapy, yet not because of severe PEG-IFN side effects in any subject. Other small trials observed SVR in 2/3 (66.6%) (129), and 2/6 (33.3%) (130) treated HCV-infected HD patients. Another retrospective study found ETR in 3/7 (42.8%) HD patients treated with PEG-IFN α -2a while the follow-up was insufficient to present SVR rate (131).

The combination of PEG-IFN α -2b and low dose ribavirin (200-400 mg/day initially) was used by Bruchfeld *et al.* (132) who reported a SVR rate of

50% (3/6), 1 death from cardiac arrest and discontinuation of treatment in 2 patients because of side effects. Despite some encouraging results obtained, since the treatment with ribavirin in this study was concentration controlled and thereby individualized and such a practice might not be easily applicable in a clinical setting, the same combination therapy cannot be recommended.

Currently, there is a clear lack of strong evidence on using PEG-IFN in HCV-infected HD patients because most of the so far published data on the topic incorporated small number of patients, were divergent in results, were not controlled, and did not compare the risk-benefit ratio of pegylated versus standard IFN formulations. Well-designed, large clinical trials are clearly required to improve our knowledge and suggest the best practice.

Summary

The prevalence of HCV infection varies greatly among various populations of patients on HD from different geographic regions. The reports since 1999 gathered in this review presented a range for HCV seroprevalence among HD patients from 1.9% in Slovenia to 80% in Senegal.

Whereas blood transfusions seems not to be a current risk factor for HCV infection in HD population, the length of time on HD was repeatedly introduced in recent reports as the major risk factor. Although the value of serology in detecting HCV infection in HD subjects is recently questioned, regarding the results of large studies, routine testing for HCV-RNA to screen HD patients for HCV infection seems unjustified. Performing ELISA for HCVcAg is a new advance in the diagnosis of HCV infection in HD patients that with its ease and low-cost in comparison to molecular studies would add the sensitivity of the screening program, especially in poor-resource settings. While mortality was found to be higher in HCV-infected than non-infected HD patients, this increment is at least partially related to chronic liver disease with its attendant complications. Since biochemical and virological testing cannot predict the extent of liver damage in HCV-infected HD persons, liver biopsy is the only accurate means of assessing the severity of the HCV infection. Strict adherence to universal precautions seems to be enough to control disease spread in HD units and isolation of HCV-infected HD patients and use of dedicated machines are currently unjustified. IFN- α monotherapy, 3 million units, subcutaneously, trice weekly for 48 weeks for all genotypes, offers a SVR rates ranged from 19% to 59% and is the only regimen that can currently be recommended.

References

- Global surveillance and control of hepatitis C. Report of a WHO Consultation organized in collaboration with the Viral Hepatitis Prevention Board, Antwerp, Belgium. *J Viral Hepat* 1999; **6**: 35-47.
- Yen T, Keeffe EB, Ahmed A. The epidemiology of hepatitis C virus infection. *J Clin Gastroenterol* 2003; **36**: 47-53.
- Strader DB, Wright T, Thomas DL, Seeff LB. Diagnosis, management, and treatment of hepatitis C. *Hepatology* 2004; **39**: 1147-71.
- Johnson RJ, Gretch DR, Yamabe H, Hart J, Bacchi CE, Hartwell P, et al. Membranoproliferative glomerulonephritis associated with hepatitis C virus infection. *N Engl J Med* 1993; **328**: 465-70.
- Fabrizi F, Poordad FF, Martin P. Hepatitis C infection and the patient with end-stage renal disease. *Hepatology* 2002; **36**: 3-10.
- Prince AM, Brotman B, Grady GF, Kuhns WJ, Hazzi C, Levine RW, et al. Long-incubation post-transfusion hepatitis without serological evidence of exposure to hepatitis-B virus. *Lancet* 1974; **2**: 241-6.
- Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M. Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science* 1989; **244**: 359-62.
- Robertson B, Myers G, Howard C, Brettin T, Bukh J, Gaschen B, et al. Classification, nomenclature, and database development for hepatitis C virus (HCV) and related viruses: proposals for standardization. International Committee on Virus Taxonomy. *Arch Virol* 1998; **143**: 2493-503.
- Pawlotsky JM. Hepatitis C virus genetic variability: pathogenic and clinical implications. *Clin Liver Dis* 2003; **7**: 45-66.
- Wreghitt TG. Blood-borne virus infections in dialysis units--a review. *Rev Med Virol* 1999; **9**: 101-9.
- Jadoul M, Poignet JL, Geddes C, Locatelli F, Medin C, Krajewska M, et al. The changing epidemiology of hepatitis C virus (HCV) infection in haemodialysis: European multicentre study. *Nephrol Dial Transplant* 2004; **19**: 904-9.
- Gallego E, Lopez A, Perez J, Llamas F, Lorenzo I, Lopez E, et al. Effect of isolation measures on the incidence and prevalence of hepatitis C virus infection in hemodialysis. *Nephron* 2006; **104**: c1-6.
- Carneiro MA, Teles SA, Dias MA, Ferreira RC, Naghettine AV, Silva SA, et al. Decline of hepatitis C infection in hemodialysis patients in Central Brazil: a ten years of surveillance. *Mem Inst Oswaldo Cruz* 2005; **100**: 345-9.
- Yang CS, Chang HH, Chou CC, Peng SJ. Isolation effectively prevents the transmission of hepatitis C virus in the hemodialysis unit. *J Formos Med Assoc* 2003; **102**: 79-85.
- Barril G, Traver JA. Decrease in the hepatitis C virus (HCV) prevalence in hemodialysis patients in Spain: effect of time, initiating HCV prevalence studies and adoption of isolation measures. *Antiviral Res* 2003; **60**: 129-34.
- Shamshirsaz AA, Kamgar M, Bekheirnia MR, Ayazi F, Hashemi SR, Bouzari N, et al. The role of hemodialysis machines dedication in reducing Hepatitis C transmission in the dialysis setting in Iran: a multicenter prospective interventional study. *BMC Nephrol* 2004; **5**: 13.
- Saxena AK, Panhotra BR, Sundaram DS, Naguib M, Venkateshappa CK, Uzzaman W, et al. Impact of dedicated space, dialysis equipment, and nursing staff on the transmission of hepatitis C virus in a hemodialysis unit of the middle east. *Am J Infect Control* 2003; **31**: 26-33.
- Valtuille R, Moretto H, Lef L, Rendo P, Fernandez JL. Decline of high hepatitis C virus prevalence in a hemodialysis unit with no isolation measures during a 6-year follow-up. *Clin Nephrol* 2002; **57**: 371-5.
- Aucella F, Vigilante M, Valente GL, Stallone C. Systematic monitor disinfection is effective in limiting HCV spread in hemodialysis. *Blood Purif* 2000; **18**: 110-4.
- Buturovic-Ponikvar J. Renal replacement therapy in Slovenia: annual report 2001. *Nephrol Dial Transplant* 2003; **18**: v53-5.
- Diouf ML, Diouf B, Niang A, Ka EH, Pouye A, Seck A, et al. Prevalence of hepatitis B and C viruses in a chronic hemodialysis center in Dakar. *Dakar Med* 2000; **45**: 1-4.
- Ahmetagic S, Muminhodzic K, Cickusic E, Stojic V, Petrovic J, Tihic N. Hepatitis C infection in risk groups. *Bosn J Basic Med Sci* 2006; **6**: 13-7.
- Salama G, Rostaing L, Sandres K, Izopet J. Hepatitis C virus infection in French hemodialysis units: a multicenter study. *J Med Virol* 2000; **61**: 44-51.
- Hinrichsen H, Leimenstoll G, Stegen G, Schrader H, Folsch UR, Schmidt WE. Prevalence and risk factors of hepatitis C virus infection in haemodialysis patients: a multicentre study in 2796 patients. *Gut* 2002; **51**: 429-33.
- Rigopoulou EI, Stefanidis I, Liaskos C, Zervou EK, Rizos C, Mina P, et al. HCV-RNA qualitative assay based on transcription mediated amplification improves the detection of hepatitis C virus infection in patients on hemodialysis: results from five hemodialysis units in central Greece. *J Clin Virol* 2005; **34**: 81-5.
- Garinis G, Spanakis N, Theodorou V, Gorgoulis V, Manolis E, Karameris A, et al. Comparison of the enzyme-linked immunosorbant assay III, recombinant immunoblot third generation assay, and polymerase chain reaction method in the detection of hepatitis C virus infection in haemodialysis patients. *J Clin Lab Anal* 1999; **13**: 122-5.
- Sypsa V, Psychogiou M, Katsoulidou A, Skoutelis G, Moutafis S, Hadjiconstantinou V, et al. Incidence and patterns of hepatitis C virus seroconversion in a cohort of hemodialysis patients. *Am J Kidney Dis* 2005; **45**: 334-43.
- Lombardi M, Cerrai T, Geatti S, Negroni S, Pertusini L, Pegoraro M, et al. Results of a national epidemiological investigation on HCV infection among dialysis patients. (Survey by the Italian Branch of EDTNA/ERCA). *J Nephrol* 1999; **12**: 322-7.
- Petrosillo N, Gilli P, Serraino D, Dentico P, Mele A, Ragni P, et al. Prevalence of infected patients and understaffing have a role in hepatitis C virus transmission in dialysis. *Am J Kidney Dis* 2001; **37**: 1004-10.
- Covic A, Iancu L, Apetrei C, Scripcaru D, Volovat C, Mititiuc I, et al. Hepatitis virus infection in haemodialysis patients from Moldavia. *Nephrol Dial Transplant* 1999; **14**: 40-5.
- Schneeberger PM, Keur I, van Loon AM, Mortier D, de Coul KO, van Haperen AV, et al. The prevalence and incidence of hepatitis C virus infections among dialysis patients in the Netherlands: a nationwide prospective study. *J Infect Dis* 2000; **182**: 1291-9.
- Almroth G, Ekerme B, Mansson AS, Svensson G, Widell A. Detection and prevention of hepatitis C in dialysis patients and renal transplant recipients. A long-term follow up (1989-January 1997). *J Intern Med* 2002; **251**: 119-28.
- Kelley VA, Everett-Kitchens J, Brannon LE, Connor K, Martinez EJ, Pearson TC, et al. Lack of seronegative hepatitis C virus infections in patients with chronic renal failure. *Transplantation* 2002; **74**: 1473-5.
- Kalantar-Zadeh K, Miller LG, Daar ES. Diagnostic discordance for hepatitis C virus infection in hemodialysis patients. *Am J Kidney Dis* 2005; **46**: 290-300.

35. Sivapalasingam S, Malak SF, Sullivan JF, Lorch J, Sepkowitz KA. High prevalence of hepatitis C infection among patients receiving hemodialysis at an urban dialysis center. *Infect Control Hosp Epidemiol* 2002; **23**: 319-24.
36. Saab S, Martin P, Brezina M, Gitnick G, Yee HF, Jr. Serum alanine aminotransferase in hepatitis c screening of patients on hemodialysis. *Am J Kidney Dis* 2001; **37**: 308-15.
37. Kalantar-Zadeh K, Kilpatrick RD, McAllister CJ, Miller LG, Daar ES, Gjertson DW, et al. Hepatitis C virus and death risk in hemodialysis patients. *J Am Soc Nephrol* 2007; **18**: 1584-93.
38. Daw MA, Elkaber MA, Drah AM, Werfalli MM, Mihat AA, Siala IM. Prevalence of Hepatitis C virus antibodies among different populations of relative and attributable risk. *Saudi Med J* 2002; **23**: 1356-60.
39. El-Amin HH, Osman EM, Mekki MO, Abdelraheem MB, Ismail MO, Yousif ME, et al. Hepatitis C virus infection in hemodialysis patients in Sudan: two centers' report. *Saudi J Kidney Dis Transpl* 2007; **18**: 101-6.
40. Bouzgarrou N, Fodha I, Othman SB, Achour A, Grattard F, Trabelsi A, et al. Evaluation of a total core antigen assay for the diagnosis of hepatitis C virus infection in hemodialysis patients. *J Med Virol* 2005; **77**: 502-8.
41. Ayed K, Gorgi Y, Ben Abdallah T, Aouadi H, Jendoubi-Ayed S, Sfar I, et al. Hepatitis C virus infection in hemodialysis patients from Tunisia: national survey by serologic and molecular methods. *Transplant Proc* 2003; **35**: 2573-5.
42. Albuquerque AC, Coelho MR, Lopes EP, Lemos MF, Moreira RC. Prevalence and risk factors of hepatitis C virus infection in hemodialysis patients from one center in Recife, Brazil. *Mem Inst Oswaldo Cruz* 2005; **100**: 467-70.
43. Carneiro MA, Martins RM, Teles SA, Silva SA, Lopes CL, Cardoso DD, et al. Hepatitis C prevalence and risk factors in hemodialysis patients in Central Brazil: a survey by polymerase chain reaction and serological methods. *Mem Inst Oswaldo Cruz* 2001; **96**: 765-9.
44. Mendez-Sanchez N, Motola-Kuba D, Chavez-Tapia NC, Bahena J, Correa-Rotter R, Uribe M. Prevalence of hepatitis C virus infection among hemodialysis patients at a tertiary-care hospital in Mexico City, Mexico. *J Clin Microbiol* 2004; **42**: 4321-2.
45. Sanchez JL, Sjogren MH, Callahan JD, Watts DM, Lucas C, Abdel-Hamid M, et al. Hepatitis C in Peru: risk factors for infection, potential iatrogenic transmission, and genotype distribution. *Am J Trop Med Hyg* 2000; **63**: 242-8.
46. Lopez-Navedo PJ, Lebron-Rivera R, Gonzalez-Trapaga J, Weber-Acevedo J, Lefevre-Ramos E, Flores-de Hostos E, et al. Prevalence of hepatitis C virus infection at three hemodialysis units in the western region of Puerto Rico. *Bol Asoc Med P R* 1999; **91**: 100-2.
47. Izopet J, Sandres-Saune K, Kamar N, Salama G, Dubois M, Pasquier C, et al. Incidence of HCV infection in French hemodialysis units: a prospective study. *J Med Virol* 2005; **77**: 70-6.
48. Fabrizi F, de Vecchi AF, Como G, Lunghi G, Martin P. De novo HCV infection among dialysis patients: a prospective study by HCV core antigen ELISA assay. *Aliment Pharmacol Ther* 2005; **21**: 861-9.
49. Fabrizi F, Martin P, Dixit V, Brezina M, Russell J, Conrad A, et al. Detection of de novo hepatitis C virus infection by polymerase chain reaction in hemodialysis patients. *Am J Nephrol* 1999; **19**: 383-8.
50. Kumagai J, Komiya Y, Tanaka J, Katayama K, Tatsukawa Y, Yorioka N, et al. Hepatitis C virus infection in 2,744 hemodialysis patients followed regularly at nine centers in Hiroshima during November 1999 through February 2003. *J Med Virol* 2005; **76**: 498-502.
51. Furusyo N, Hayashi J, Kakuda K, Ariyama I, Kanamoto-Tanaka Y, Shimizu C, et al. Acute hepatitis C among Japanese hemodialysis patients: a prospective 9-year study. *Am J Gastroenterol* 2001; **96**: 1592-600.
52. Fissell RB, Bragg-Gresham JL, Woods JD, Jadoul M, Gillespie B, Hedderwick SA, et al. Patterns of hepatitis C prevalence and seroconversion in hemodialysis units from three continents: the DOPPS. *Kidney Int* 2004; **65**: 2335-42.
53. Moreira R, Pinho JR, Fares J, Oba IT, Cardoso MR, Saraceni CP, et al. Prospective study of hepatitis C virus infection in hemodialysis patients by monthly analysis of HCV RNA and antibodies. *Can J Microbiol* 2003; **49**: 503-7.
54. Santos MA, Souto FJ. Infection by the hepatitis C virus in chronic renal failure patients undergoing hemodialysis in Mato Grosso state, central Brazil: a cohort study. *BMC Public Health* 2007; **7**: 32.
55. Amiri ZM, Shakib AJ, Toorchi M. Seroprevalence of hepatitis C and risk factors in haemodialysis patients in Guilan, Islamic Republic of Iran. *East Mediterr Health J* 2005; **11**: 372-6.
56. Alavian SM, Einollahi B, Hajarizadeh B, Bakhtiari S, Nafar M, Ahrabi S. Prevalence of hepatitis C virus infection and related risk factors among Iranian haemodialysis patients. *Nephrology* 2003; **8**: 256-60.
57. Ansari MM, Kooloobandi A. Prevalence of hepatitis C virus infection in thalassemia and haemodialysis patients in north Iran-Rasht. *J Viral Hepat* 2002; **9**: 390-2.
58. Bdour S. Hepatitis C virus infection in Jordanian haemodialysis units: serological diagnosis and genotyping. *J Med Microbiol* 2002; **51**: 700-4.
59. Hussein MM, Mooij JM, Hegazy MS, Bamaga MS. The impact of polymerase chain reaction assays for the detection of hepatitis C virus infection in a hemodialysis unit. *Saudi J Kidney Dis Transpl* 2007; **18**: 107-13.
60. Al-Shohaib S, Abd-Elaal M, Zawawi T, Abbas F, Shaheen F, Amoah E. The prevalence of hepatitis C virus antibodies among hemodialysis patients in Jeddah area, Saudi Arabia. *Saudi Med J* 2003; **2**: S125.
61. Ben Othman S, Bouzgarrou N, Achour A, Bourlet T, Pozzetto B, Trabelsi A. High prevalence and incidence of hepatitis C virus infections among dialysis patients in the East-Centre of Tunisia. *Pathol Biol* 2004; **52**: 323-7.
62. Othman B, Monem F. Prevalence of antibodies to hepatitis C virus among hemodialysis patients in Damascus, Syria. *Infection* 2001; **29**: 262-5.
63. Lopez-Alcorocho JM, Barril G, Ortiz-Movilla N, Traver JA, Bartolome J, Sanz P, et al. Prevalence of hepatitis B, hepatitis C, GB virus C/hepatitis G and TT viruses in predialysis and hemodialysis patients. *J Med Virol* 2001; **63**: 103-7.
64. Hosseini-Moghaddam SM, Keyvani H, Kasiri H, Kazemeyni SM, Basiri A, Aghel N, et al. Distribution of hepatitis C virus genotypes among hemodialysis patients in Tehran--a multicenter study. *J Med Virol* 2006; **78**: 569-73.
65. Ocak S, Duran N, Kaya H, Emir I. Seroprevalence of hepatitis C in patients with type 2 diabetes mellitus and non-diabetic on haemodialysis. *Int J Clin Pract* 2006; **60**: 670-4.
66. Rivanera D, Lilli D, Lorino G, Pirozzi V, Cannulla V, Dicunzio G, et al. Detection of antibodies to hepatitis C virus in dialysis patients. *Eur J Epidemiol* 1993; **9**: 55-8.
67. McHutchison JG, Person JL, Govindarajan S, Valinluck B, Gore T, Lee SR, et al. Improved detection of hepatitis C virus antibodies in high-risk populations. *Hepatology* 1992; **15**: 19-25.
68. Chaudhary RK, Andonov A, MacLean C. Detection of

- hepatitis C virus infection with recombinant immunoblot assay, synthetic immunoblot assay, and polymerase chain reaction. *J Clin Lab Anal* 1993; **7**: 164-7.
69. Saab S, Brezina M, Gitnick G, Martin P, Yee HF, Jr. Hepatitis C screening strategies in hemodialysis patients. *Am J Kidney Dis* 2001; **38**: 91-7.
 70. Neng Lai K. Hepatitis C infection screening in hemodialysis units. *Am J Kidney Dis* 2001; **38**: 186-8.
 71. Scott JD, Gretch DR. Molecular diagnostics of hepatitis C virus infection: a systematic review. *JAMA* 2007; **297**: 724-32.
 72. Recommendations for prevention and control of hepatitis C virus (HCV) infection and HCV-related chronic disease. Centers for Disease Control and Prevention. *MMWR Recomm Rep* 1998; **47**: 1-39.
 73. Recommendations for preventing transmission of infections among chronic hemodialysis patients. *MMWR Recomm Rep* 2001; **50**: 1-43.
 74. Fabrizi F, Lunghi G, Finazzi S, Colucci P, Pagano A, Ponticelli C, et al. Decreased serum aminotransferase activity in patients with chronic renal failure: impact on the detection of viral hepatitis. *Am J Kidney Dis* 2001; **38**: 1009-15.
 75. Khaja MN, Madhavi C, Thippavazzula R, Nafeesa F, Habib AM, Habibullah CM, et al. High prevalence of hepatitis C virus infection and genotype distribution among general population, blood donors and risk groups. *Infect Genet Evol* 2006; **6**: 198-204.
 76. Sullivan DG, Kim SS, Wilson JJ, Stehman-Breen C, Gretch DR. Investigating hepatitis C virus heterogeneity in a high prevalence setting using heteroduplex tracking analysis. *J Virol Methods* 2001; **96**: 5-16.
 77. Courouce AM, Le Marrec N, Bouchardeau F, Razer A, Maniez M, Laperche S, et al. Efficacy of HCV core antigen detection during the preseroconversion period. *Transfusion* 2000; **40**: 1198-202.
 78. Aoyagi K, Iida K, Ohue C, Matsunaga Y, Tanaka E, Kiyosawa K, et al. Performance of a conventional enzyme immunoassay for hepatitis C virus core antigen in the early phases of hepatitis C infection. *Clin Lab* 2001; **47**: 119-27.
 79. Martin P, Carter D, Fabrizi F, Dixit V, Conrad AJ, Artinian L, et al. Histopathological features of hepatitis C in renal transplant candidates. *Transplantation* 2000; **69**: 1479-84.
 80. Sterling RK, Sanyal AJ, Luketic VA, Stravitz RT, King AL, Post AB, et al. Chronic hepatitis C infection in patients with end stage renal disease: characterization of liver histology and viral load in patients awaiting renal transplantation. *Am J Gastroenterol* 1999; **94**: 3576-82.
 81. Cotler SJ, Diaz G, Gundlapalli S, Jakate S, Chawla A, Mital D, et al. Characteristics of hepatitis C in renal transplant candidates. *J Clin Gastroenterol* 2002; **35**: 191-5.
 82. Sezer S, Ozdemir BH, Arat Z, Turan M, Ozdemir NF, Haberal M. Spectrum of liver damage and correlation with clinical and laboratory parameters in HCV infected hemodialysis patients. *Ren Fail* 2001; **23**: 807-18.
 83. Boyacioglu S, Gur G, Yilmaz U, Korkmaz M, Demirhan B, Bilezikci B, et al. Investigation of possible clinical and laboratory predictors of liver fibrosis in hemodialysis patients infected with hepatitis C virus. *Transplant Proc* 2004; **36**: 50-2.
 84. Furusyo N, Hayashi J, Kanamoto-Tanaka Y, Ariyama I, Etoh Y, Shigematsu M, et al. Liver damage in hemodialysis patients with hepatitis C virus viremia: a prospective 10-year study. *Dig Dis Sci* 2000; **45**: 2221-8.
 85. Meyers CM, Seeff LB, Stehman-Breen CO, Hoofnagle JH. Hepatitis C and renal disease: an update. *Am J Kidney Dis* 2003; **42**: 631-57.
 86. Rampino T, Arbustini E, Gregorini M, Guallini P, Libetta C, Maggio M, et al. Hemodialysis prevents liver disease caused by hepatitis C virus: role of hepatocyte growth factor. *Kidney Int* 1999; **56**: 2286-91.
 87. Akpolat I, Ozyilkan E, Karagoz F, Akpolat T, Kandemir B. Hepatitis C in haemodialysis and nonuraemic patients: a histopathological study. *Int Urol Nephrol* 1998; **30**: 349-55.
 88. Luzar B, Ferlan-Marolt V, Brinovec V, Lesnicar G, Klopčič U, Poljak M. Does end-stage kidney failure influence hepatitis C progression in hemodialysis patients? *Hepatogastroenterology* 2003; **50**: 157-60.
 89. Fabrizi F, Martin P, Dixit V, Brezina M, Cole MJ, Gerosa S, et al. Quantitative assessment of HCV load in chronic hemodialysis patients: a cross-sectional survey. *Nephron* 1998; **80**: 428-33.
 90. Badalamenti S, Catania A, Lunghi G, Covini G, Bredi E, Brancaccio D, et al. Changes in viremia and circulating interferon-alpha during hemodialysis in hepatitis C virus-positive patients: only coincidental phenomena? *Am J Kidney Dis* 2003; **42**: 143-50.
 91. Nakayama E, Akiba T, Marumo F, Sato C. Prognosis of anti-hepatitis C virus antibody-positive patients on regular hemodialysis therapy. *J Am Soc Nephrol* 2000; **11**: 1896-902.
 92. Fabrizi F, Martin P, Dixit V, Bunnapradist S, Dulai G. Meta-analysis: Effect of hepatitis C virus infection on mortality in dialysis. *Aliment Pharmacol Ther* 2004; **20**: 1271-7.
 93. Iwasaki Y, Esumi M, Hosokawa N, Yanai M, Kawano K. Occasional infection of hepatitis C virus occurring in haemodialysis units identified by serial monitoring of the virus infection. *J Hosp Infect* 2000; **45**: 54-61.
 94. Kondili LA, Genovese D, Argentini C, Chionne P, Toscani P, Fabro R, et al. Nosocomial transmission in simultaneous outbreaks of hepatitis C and B virus infections in a hemodialysis center. *Eur J Clin Microbiol Infect Dis* 2006; **25**: 527-31.
 95. Halfon P, Roubicek C, Gerolami V, Quentin Y, Khiri H, Pepe G, et al. Use of phylogenetic analysis of hepatitis C virus (HCV) hypervariable region 1 sequences to trace an outbreak of HCV in an autodialysis unit. *J Clin Microbiol* 2002; **40**: 1541-5.
 96. Kalinina O, Norder H, Vetrov T, Zhdanov K, Barzunova M, Plotnikova V, et al. Shift in predominating subtype of HCV from 1b to 3a in St. Petersburg mediated by increase in injecting drug use. *J Med Virol* 2001; **65**: 517-24.
 97. Savey A, Simon F, Izopet J, Lepoutre A, Fabry J, Desenclos JC. A large nosocomial outbreak of hepatitis C virus infections at a hemodialysis center. *Infect Control Hosp Epidemiol* 2005; **26**: 752-60.
 98. Delarocque-Astagneau E, Baffoy N, Thiers V, Simon N, de Valk H, Laperche S, et al. Outbreak of hepatitis C virus infection in a hemodialysis unit: potential transmission by the hemodialysis machine? *Infect Control Hosp Epidemiol* 2002; **23**: 328-34.
 99. Katsoulidou A, Paraskevis D, Kalapothaki V, Arvanitis D, Karayiannis P, Hadjiconstantiou V, et al. Molecular epidemiology of a hepatitis C virus outbreak in a haemodialysis unit. Multicentre Haemodialysis Cohort Study on Viral Hepatitis. *Nephrol Dial Transplant* 1999; **14**: 1188-94.
 100. Jadoul M, Cornu C, van Ypersele de Strihou C. Universal precautions prevent hepatitis C virus transmission: a 54 month follow-up of the Belgian Multicenter Study. The Universitaires Cliniques St-Luc (UCL) Collaborative Group. *Kidney Int* 1998; **53**: 1022-5.
 101. Alfurayh O, Sabeel A, Al Ahdal MN, Almehari K, Kessie G, Hamid M, et al. Hand contamination with hepatitis C virus in staff looking after hepatitis C-positive hemodialysis patients. *Am J Nephrol* 2000; **20**: 103-6.

102. Harmankaya O, Cetin B, Obek A, Seber E. Low prevalence of hepatitis C virus infection in hemodialysis units: effect of isolation? *Ren Fail* 2002; **24**: 639-44.
103. Taskapan H, Oymak O, Dogukan A, Utas C. Patient to patient transmission of hepatitis C virus in hemodialysis units. *Clin Nephrol* 2001; **55**: 477-81.
104. Gilli P, Soffritti S, De Paoli Vitali E, Bedani PL. Prevention of hepatitis C virus in dialysis units. *Nephron* 1995; **70**: 301-6.
105. Froio N, Nicastrì E, Comandini UV, Cherubini C, Felicioni R, Solmone M, et al. Contamination by hepatitis B and C viruses in the dialysis setting. *Am J Kidney Dis* 2003; **42**: 546-50.
106. Canavese C, Hollo Z, Ciccone G, Ghisetti V, Barbui A, Fop F, et al. Extrahepatic immunological manifestations of hepatitis C virus in dialysis patients. *J Nephrol* 2000; **13**: 352-9.
107. Poynard T, Marcellin P, Lee SS, Niederau C, Minuk GS, Ideo G, et al. Randomised trial of interferon alpha2b plus ribavirin for 48 weeks or for 24 weeks versus interferon alpha2b plus placebo for 48 weeks for treatment of chronic infection with hepatitis C virus. International Hepatitis Interventional Therapy Group (IHIT). *Lancet* 1998; **352**: 1426-32.
108. Yee HS, Currie SL, Darling JM, Wright TL. Management and treatment of hepatitis C viral infection: recommendations from the Department of Veterans Affairs Hepatitis C Resource Center program and the National Hepatitis C Program office. *Am J Gastroenterol* 2006; **101**: 2360-78.
109. Glue P, Fang JW, Rouzier-Panis R, Raffanel C, Sabo R, Gupta SK, et al. Pegylated interferon-alpha2b: pharmacokinetics, pharmacodynamics, safety, and preliminary efficacy data. Hepatitis C Intervention Therapy Group. *Clin Pharmacol Ther* 2000; **68**: 556-67.
110. Manns MP, McHutchison JG, Gordon SC, Rustgi VK, Shiffman M, Reindollar R, et al. Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: a randomised trial. *Lancet* 2001; **358**: 958-65.
111. Hadziyannis SJ, Sette H, Jr., Morgan TR, Balan V, Diago M, Marcellin P, et al. Peginterferon-alpha2a and ribavirin combination therapy in chronic hepatitis C: a randomized study of treatment duration and ribavirin dose. *Ann Intern Med* 2004; **140**: 346-55.
112. Fried MW, Shiffman ML, Reddy KR, Smith C, Marinos G, Goncales FL, Jr., et al. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N Engl J Med* 2002; **347**: 975-82.
113. Magnone M, Holley JL, Shapiro R, Scantlebury V, McCauley J, Jordan M, et al. Interferon-alpha-induced acute renal allograft rejection. *Transplantation* 1995; **59**: 1068-70.
114. Rostaing L, Izopet J, Baron E, Duffaut M, Puel J, Durand D. Treatment of chronic hepatitis C with recombinant interferon alpha in kidney transplant recipients. *Transplantation* 1995; **59**: 1426-31.
115. Kamar N, Ribes D, Izopet J, Rostaing L. Treatment of hepatitis C virus infection (HCV) after renal transplantation: implications for HCV-positive dialysis patients awaiting a kidney transplant. *Transplantation* 2006; **82**: 853-6.
116. Russo MW, Goldsweig CD, Jacobson IM, Brown RS, Jr. Interferon monotherapy for dialysis patients with chronic hepatitis C: an analysis of the literature on efficacy and safety. *Am J Gastroenterol* 2003; **98**: 1610-5.
117. Fabrizi F, Dulai G, Dixit V, Bunnapradist S, Martin P. Meta-analysis: interferon for the treatment of chronic hepatitis C in dialysis patients. *Aliment Pharmacol Ther* 2003; **18**: 1071-81.
118. Rostaing L, Chatelut E, Payen JL, Izopet J, Thalamas C, Ton-That H, et al. Pharmacokinetics of alphaIFN-2b in chronic hepatitis C virus patients undergoing chronic hemodialysis or with normal renal function: clinical implications. *J Am Soc Nephrol* 1998; **9**: 2344-8.
119. Kamar N, Toupance O, Buchler M, Sandres-Saune K, Izopet J, Durand D, et al. Evidence that clearance of hepatitis C virus RNA after alpha-interferon therapy in dialysis patients is sustained after renal transplantation. *J Am Soc Nephrol* 2003; **14**: 2092-8.
120. Campistol JM, Esforzado N, Martinez J, Rosello L, Veciana L, Modol J, et al. Efficacy and tolerance of interferon-alpha(2b) in the treatment of chronic hepatitis C virus infection in haemodialysis patients. Pre- and post-renal transplantation assessment. *Nephrol Dial Transplant* 1999; **14**: 2704-9.
121. Huraib S, Tanimu D, Romeh SA, Quadri K, Al Ghamdi G, Iqbal A, et al. Interferon-alpha in chronic hepatitis C infection in dialysis patients. *Am J Kidney Dis* 1999; **34**: 55-60.
122. Casanovas Taltavull T, Baliellas C, Sese E, Iborra MJ, Benasco C, Andres E, et al. Interferon may be useful in hemodialysis patients with hepatitis C virus chronic infection who are candidates for kidney transplant. *Transplant Proc* 1995; **27**: 2229-30.
123. Tan AC, Brouwer JT, Glue P, van Leusen R, Kauffmann RH, Schalm SW, et al. Safety of interferon and ribavirin therapy in haemodialysis patients with chronic hepatitis C: results of a pilot study. *Nephrol Dial Transplant* 2001; **16**: 193-5.
124. Lamb M, Marks I, Wynohradnyk L. 40 kDa peginterferon alfa-2a (Pegasys) can be administered safely in patients with end-stage renal disease. *Hepatology* 2001; **34**: 326A.
125. Covic A, Maftai ID, Mardare NG, Ionita-Radu F, Totolici C, Tuta L, et al. Analysis of safety and efficacy of pegylated-interferon alpha-2a in hepatitis C virus positive hemodialysis patients: results from a large, multicenter audit. *J Nephrol* 2006; **19**: 794-801.
126. Russo MW, Ghalib R, Sigal S, Joshi V. Randomized trial of pegylated interferon alpha-2b monotherapy in haemodialysis patients with chronic hepatitis C. *Nephrol Dial Transplant* 2006; **21**: 437-43.
127. Kokoglu OF, Ucmak H, Hosoglu S, Cetinkaya A, Kantarceken B, Buyukbese MA, et al. Efficacy and tolerability of pegylated-interferon alpha-2a in hemodialysis patients with chronic hepatitis C. *J Gastroenterol Hepatol* 2006; **21**: 575-80.
128. Sporea I, Popescu A, Sirlu R, Golea O, Totolici C, Danila M, et al. Pegylated-interferon alpha 2a treatment for chronic hepatitis C in patients on chronic haemodialysis. *World J Gastroenterol* 2006; **12**: 4191-4.
129. Teta D, Luscher BL, Gonvers JJ, Francioli P, Phan O, Burnier M. Pegylated interferon for the treatment of hepatitis C virus in haemodialysis patients. *Nephrol Dial Transplant* 2005; **20**: 991-3.
130. Chan TM, Ho SK, Tang CS, Tse KC, Lam MF, Lai KN, et al. Pilot study of pegylated interferon-alpha 2a in dialysis patients with chronic hepatitis C virus infection. *Nephrology* 2007; **12**: 11-7.
131. Rivera M, Gentil MA, Sayago M, Gonzalez Roncero F, Trigo C, Algarra G, et al. Treatment of hepatitis C virus with interferon in hemodialysis patients awaiting kidney transplant. *Transplant Proc* 2005; **37**: 1424-5.
132. Bruchfeld A, Lindahl K, Reichard O, Carlsson T, Schvarcz R. Pegylated interferon and ribavirin treatment for hepatitis C in haemodialysis patients. *J Viral Hepat* 2006; **13**: 316-21.